

# Application of OMICS technologies on Gamete and Embryo Selection

**Denny Sakkas, Ph.D.**

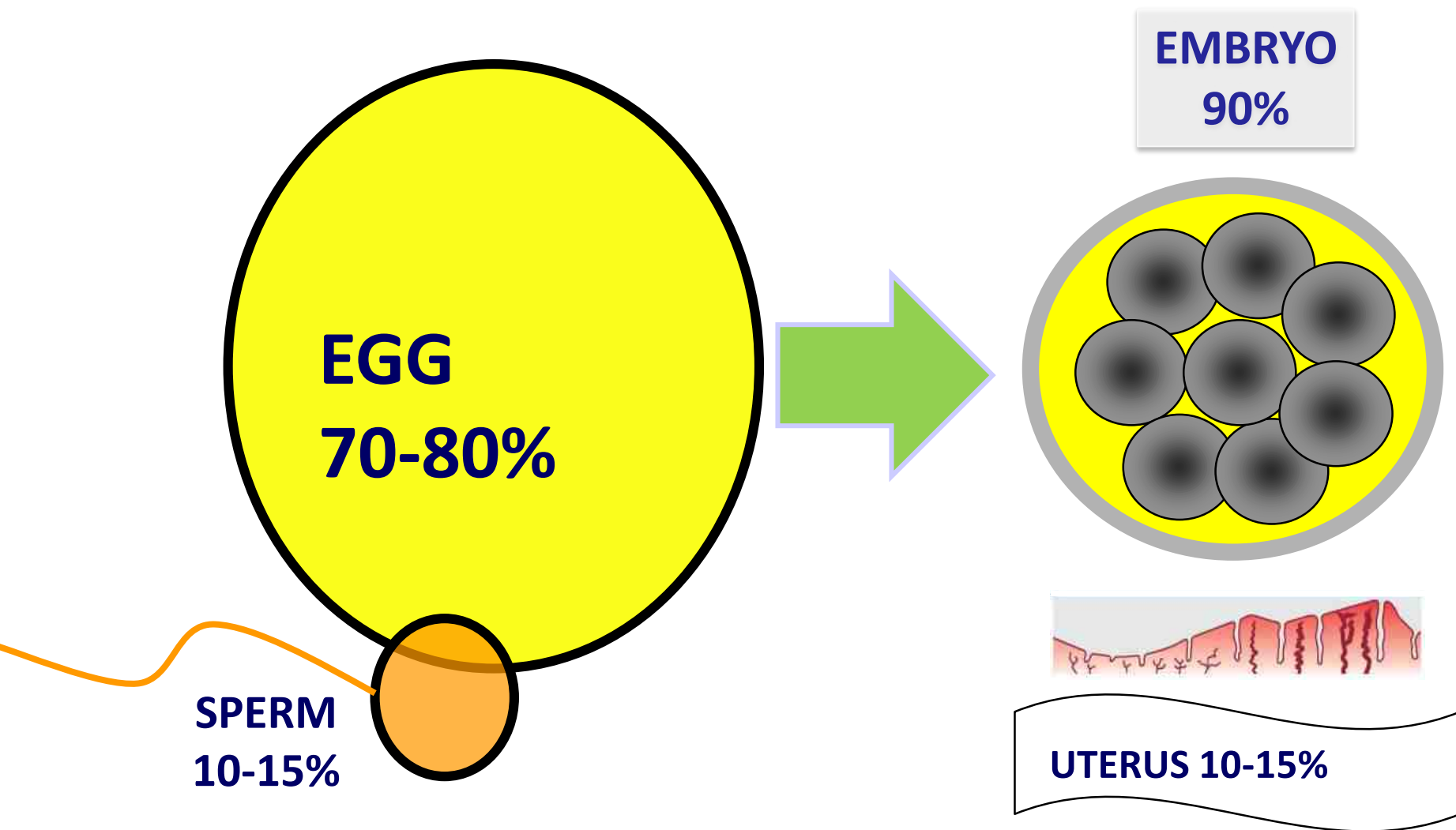
**Scientific Director,  
Boston IVF  
Waltham, MA, USA**



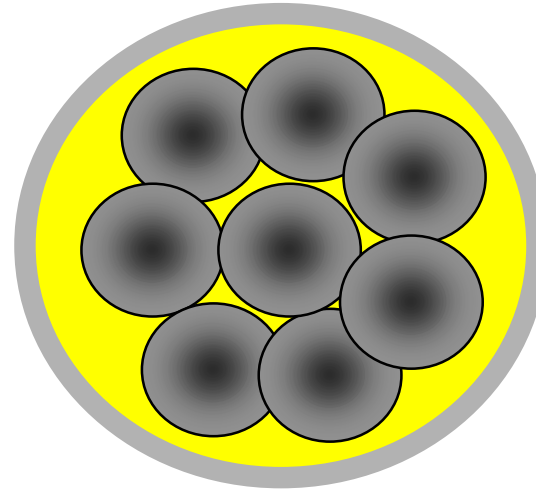
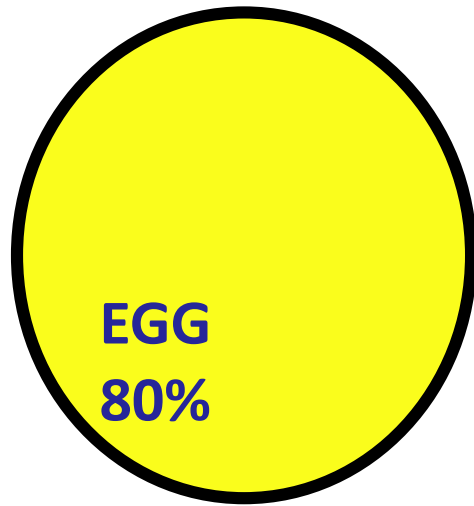
# **THE FUTURE ROLE OF THE EMBRYOLOGIST WILL FOCUS ON PROVIDING OUR PATIENTS**

- HIGHER SUCCESS RATES**
- MORE PERSONALISED DIAGNOSIS AND TREATMENT**
- AND ONE HEALTHY BABY AT A TIME**

# Venn diagram of the Responsibilities of Implantation Failure



# Venn diagram of the Responsibilities of Implantation Failure



How can the egg and embryo impact the success of Reproduction?

## Outcomes of ART Cycles Using Fresh Nondonor Eggs or Embryos, by Stage, 2012

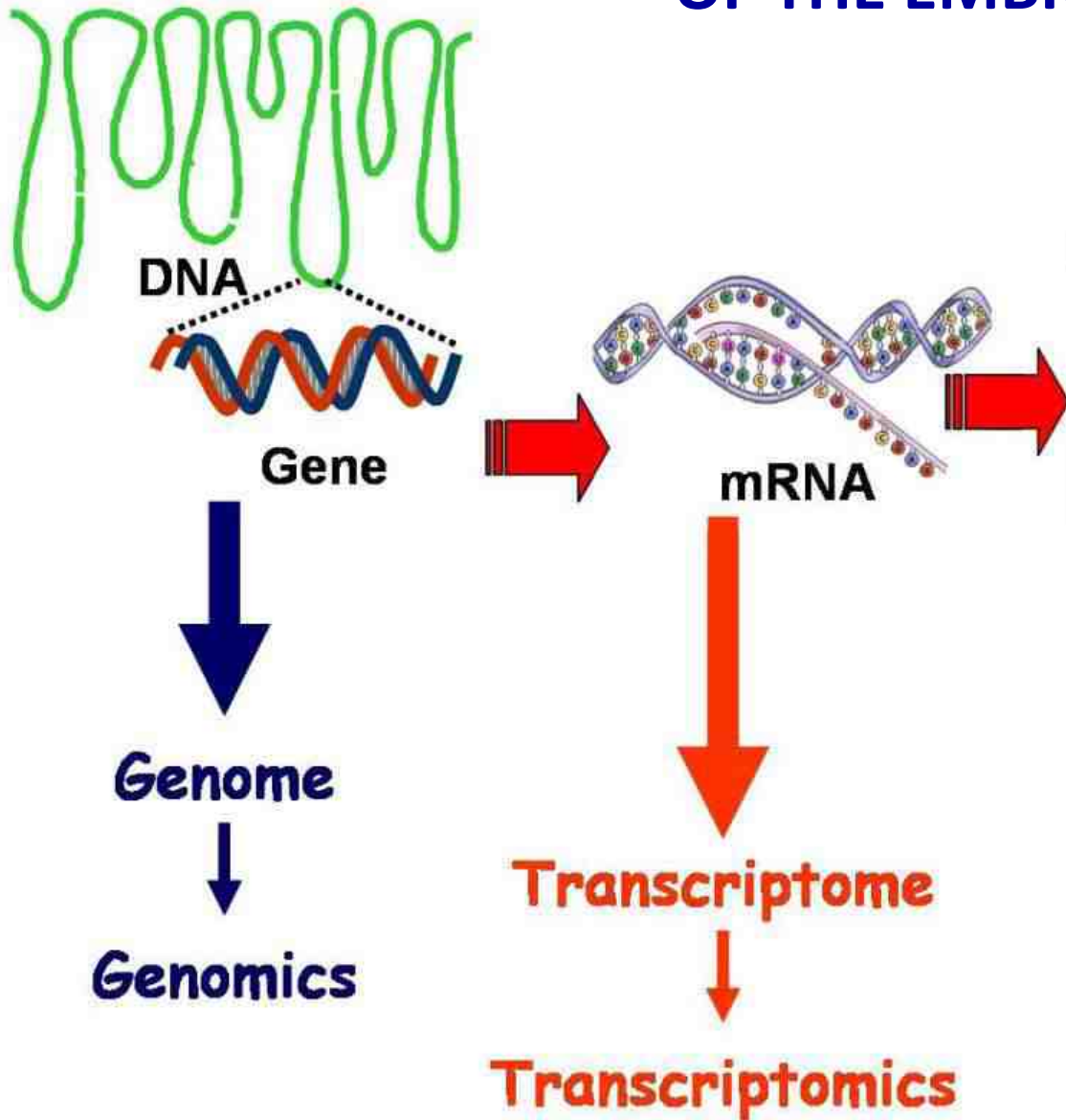


***Greatest  
Impact  
On Success***

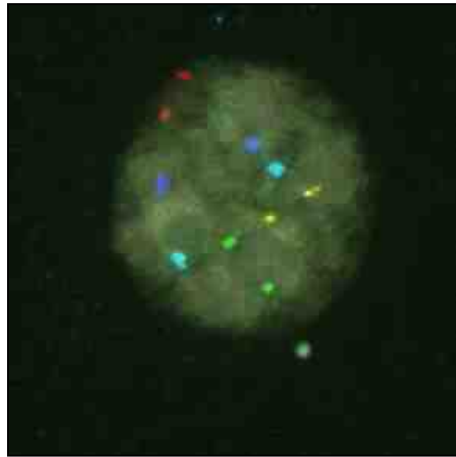
***56%***

***18%***

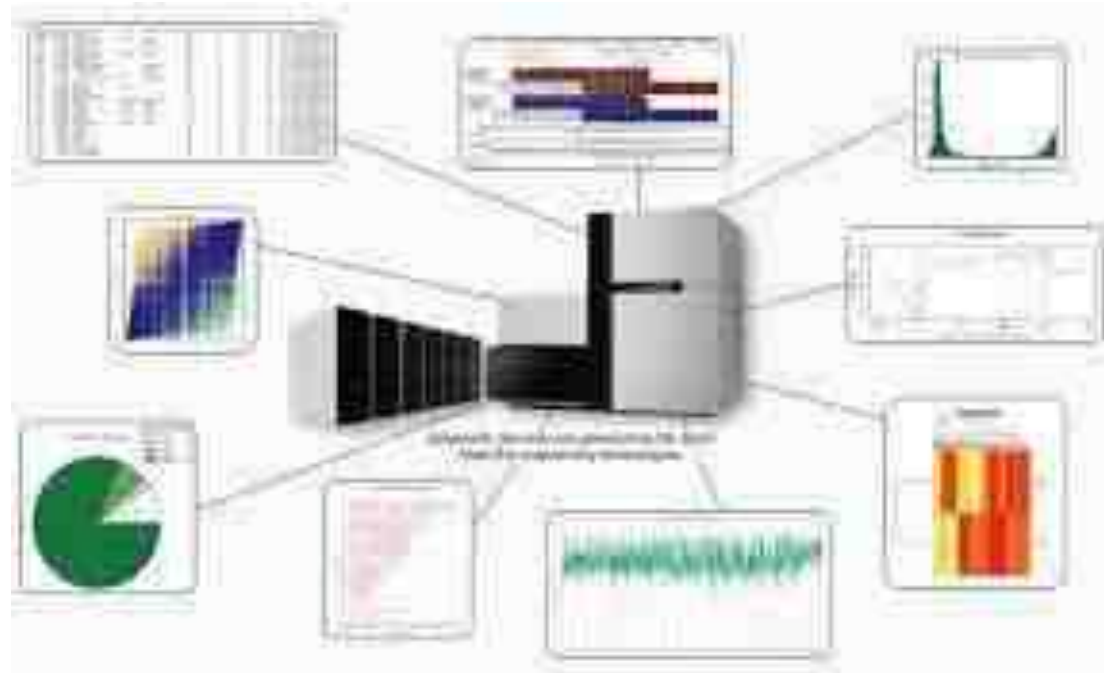
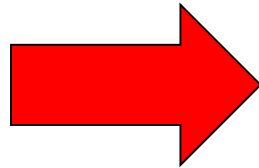
# THE “OMICS” – INVASIVE ASSESSMENT OF THE EMBRYO



# The Preimplantation Genetic Screening Example: A Strong Hypothesis Can Be A Slave To Technology

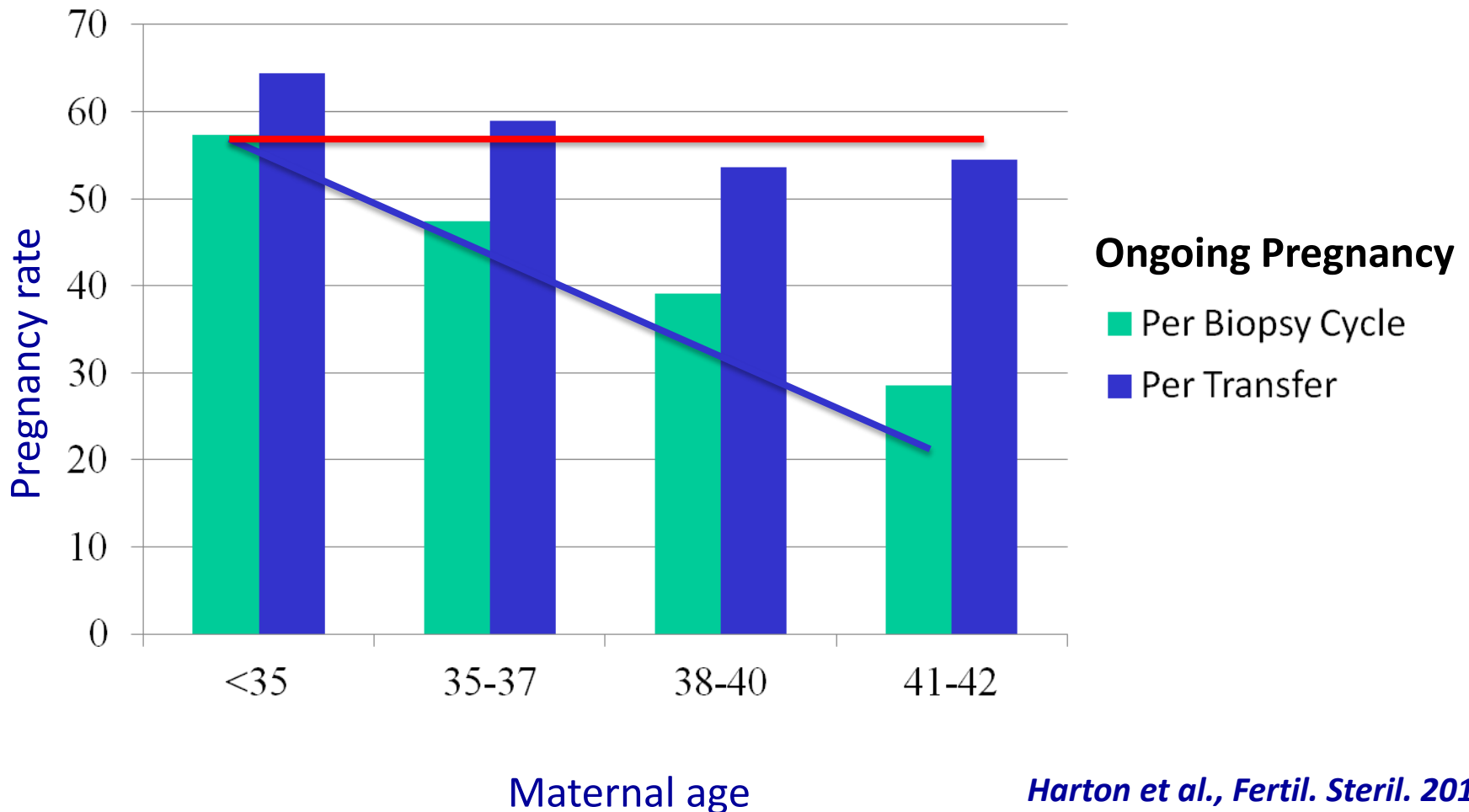


FISH  
POOR RESULTS



Array CGH or NGS  
GOOD RESULTS

# Blastocyst Biopsy and aCGH does appear to limit the effect of age when performed on Day 5



# Single embryo transfer with comprehensive chromosome screening results in improved ongoing pregnancy rates and decreased miscarriage rates

E.J. Forman<sup>1,2,\*</sup>, X. Tao<sup>1</sup>, K.M. Ferry<sup>1</sup>, D. Taylor<sup>1,2,3</sup>, N.R. Treff<sup>1,2,3</sup>, and R.T. Scott Jr<sup>1,2</sup>

**Table III** Pregnancy outcomes by treatment group.

	Control SET	CCS-SET
Chemical pregnancy rate	108/182 (59.3%)	96/140 (68.6%)
Ongoing pregnancy rate	76/182 (41.8%)	77/140 (55.0%)
Clinical miscarriage rate	25/101 (24.8%)	9/86 (10.5%)
Monozygotic twin rate	0/76 (0%)	1/77 (1.3%)
Gestational age at delivery (weeks ± SD)	38.9 ± 1.4	38.6 ± 2.0
Birthweight (grams ± SD)	3286 ± 522	3281 ± 525

\*Chi-square.

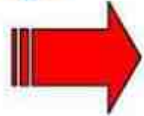
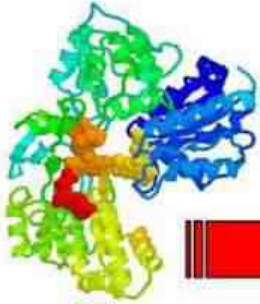
†Fisher's exact test.

‡t-test.

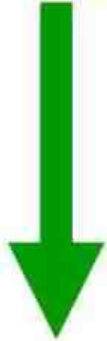
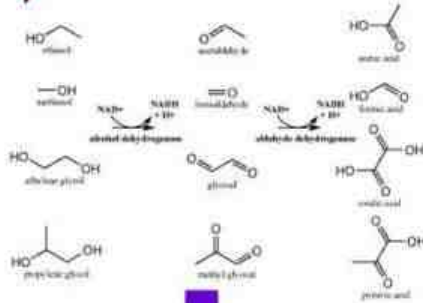
# THE OMICS – NON-INVASIVE ASSESSMENT OF THE EMBRYO



**Proteins**



**Metabolites**

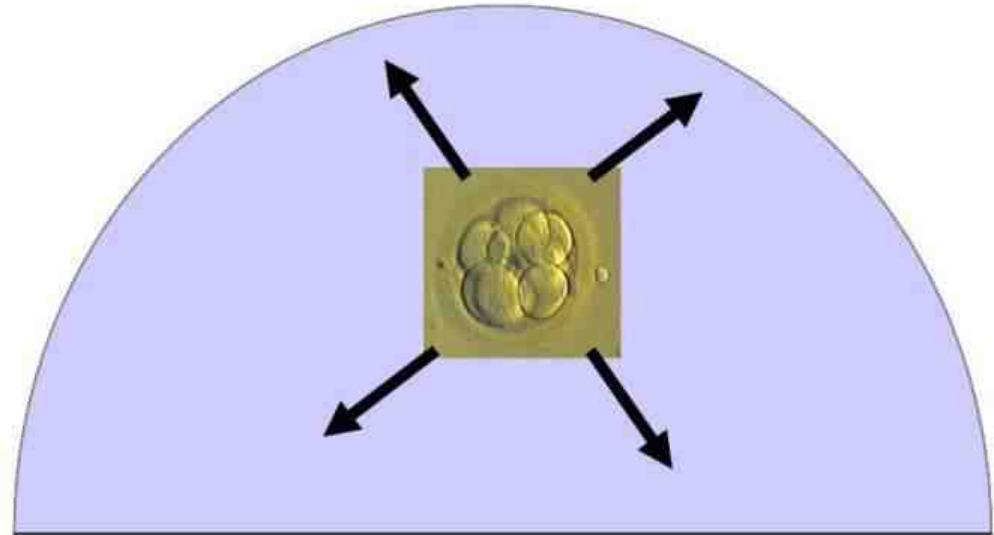


**Proteome**



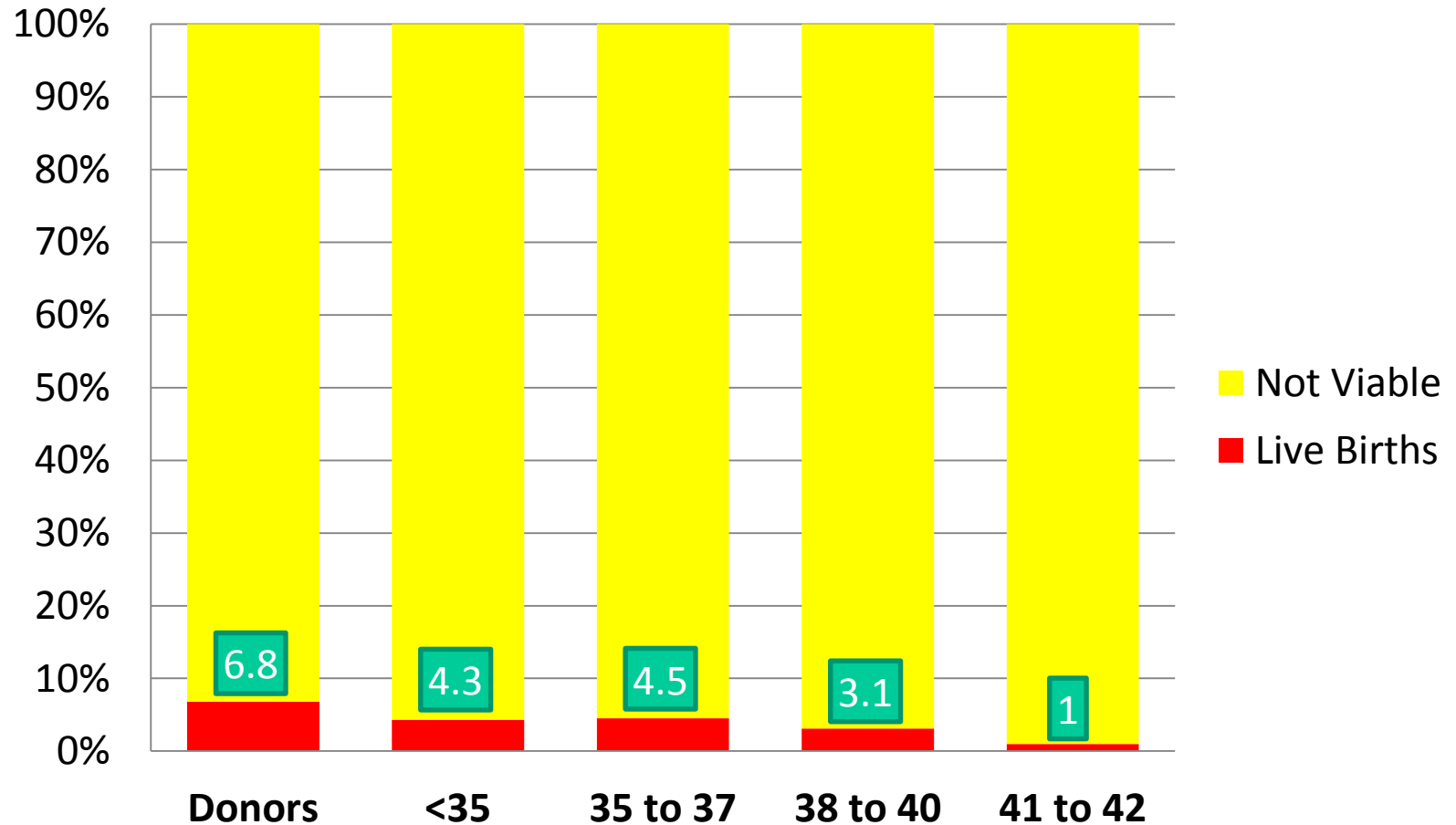
**Metabolome**

**NON-INVASIVE**

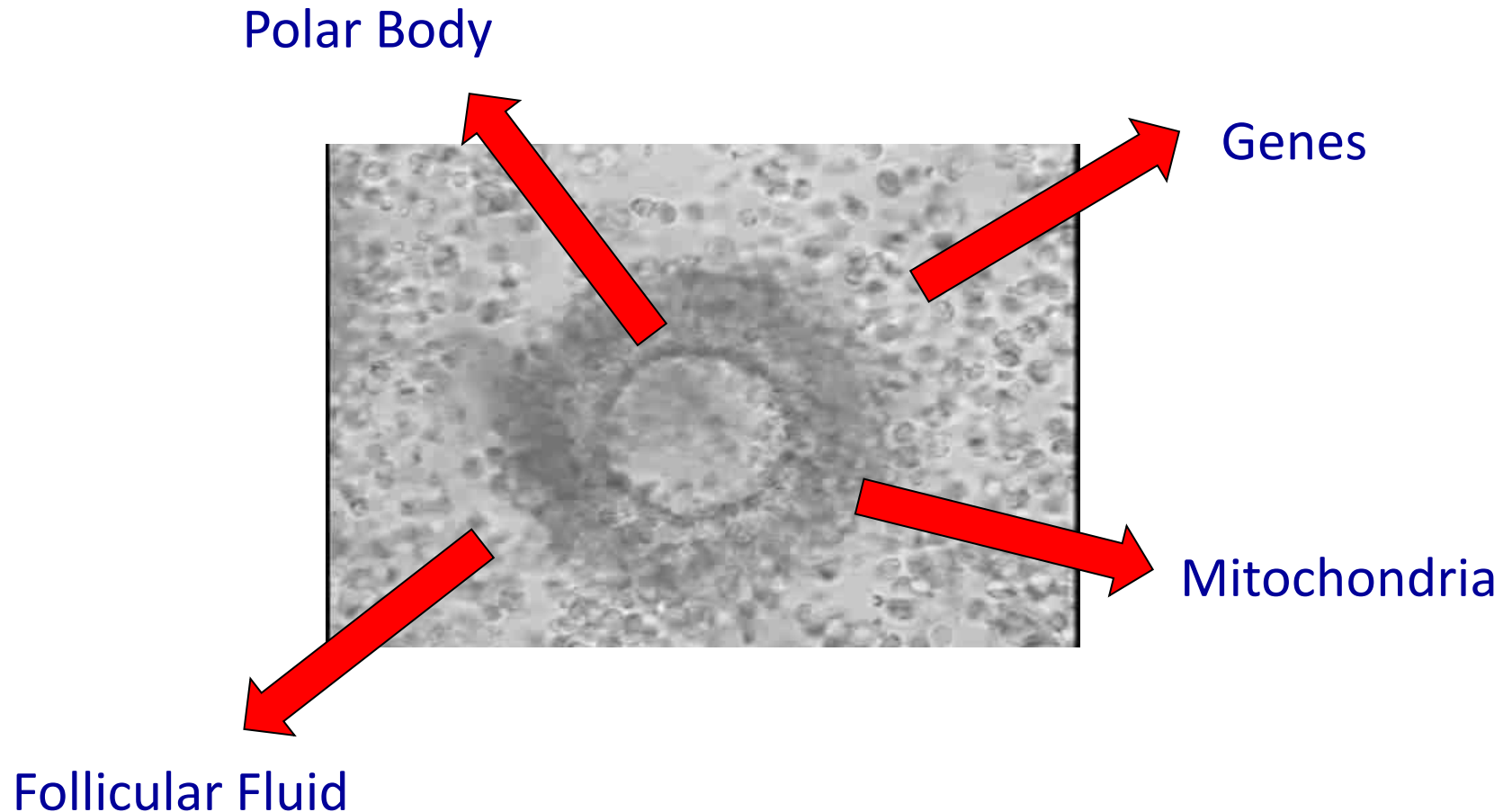


# How many live births occur per 100 oocytes retrieved?

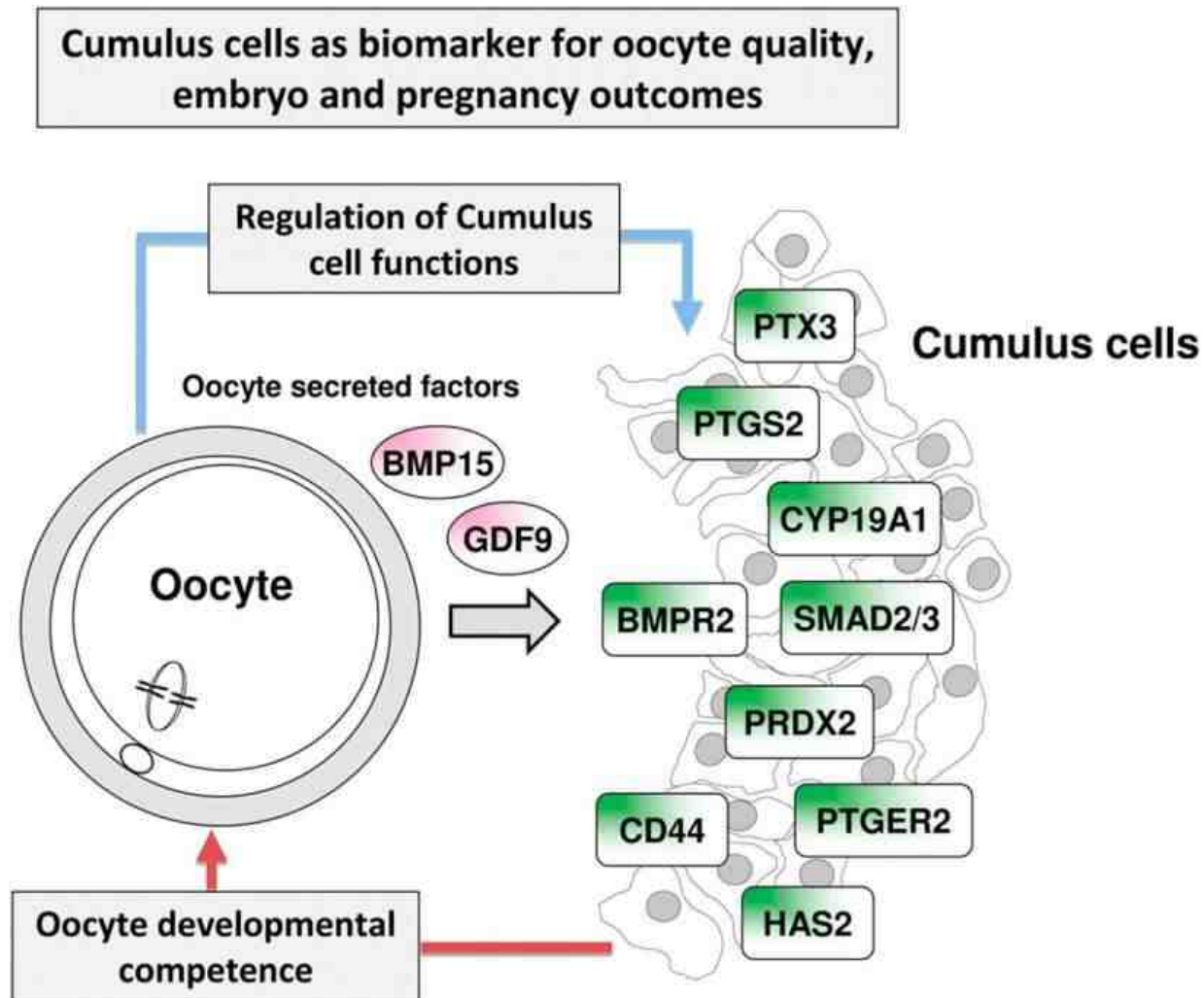
## Live Births Per Oocyte Retrieved



# Cumulus or Follicular cells as surrogate markers of viability



# Potential role of CCs as a regulator of oocyte competence and as biomarkers for oocyte/embryo quality or pregnancy outcome.



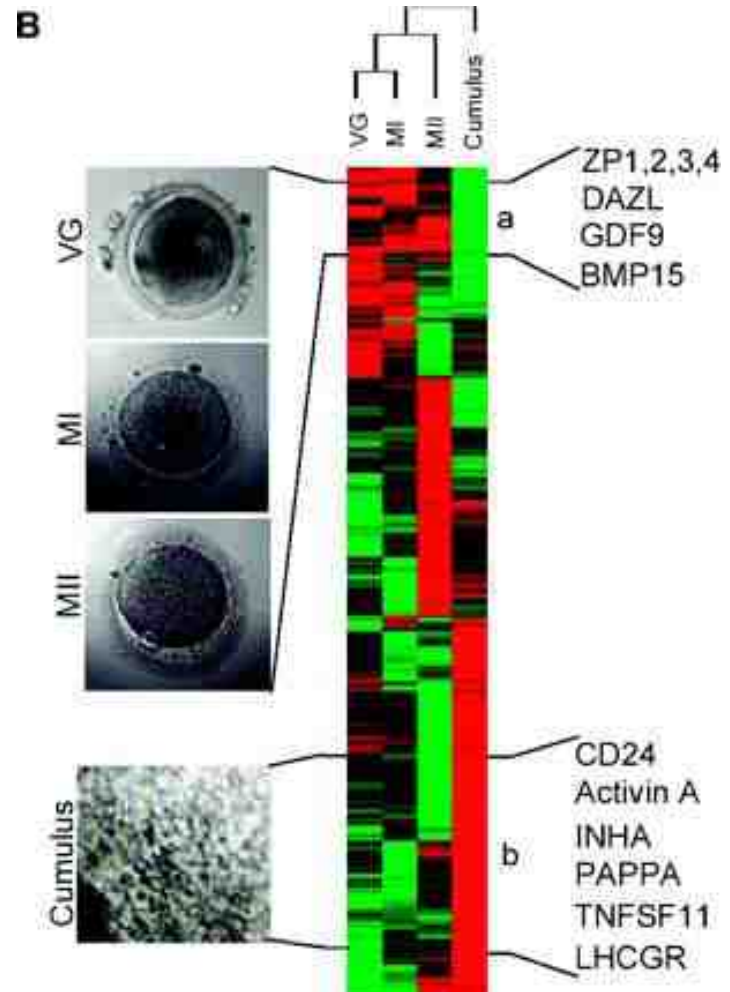
Assou S et al. Mol. Hum. Reprod. 2010;16:531-538

# Cumulus or Follicular cells as surrogate markers of viability

Hamamah et al. Comparative protein expression profiling in human cumulus cells in relation to oocyte fertilization and ovarian stimulation protocol. *Reprod Biomed Online*. 2006 13:807-14.

Feuerstein et al. Gene expression in human cumulus cells: one approach to oocyte competence. *Hum Reprod*. 2007 22:3069-77.

Hamel et al. Identification of differentially expressed markers in human follicular cells associated with competent oocytes. *Hum Reprod*. 2008 23:1118-27.

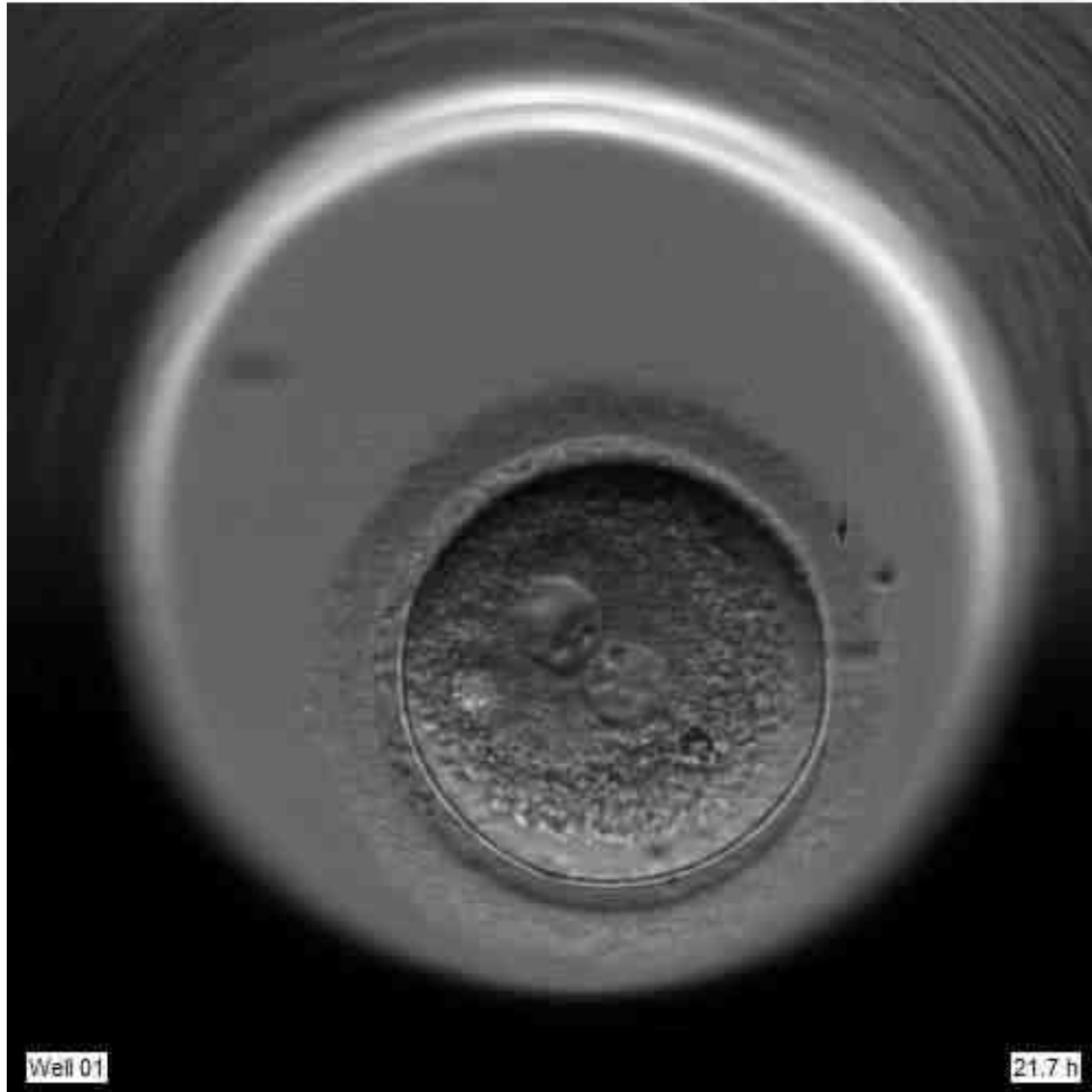


# Real Time Morphology



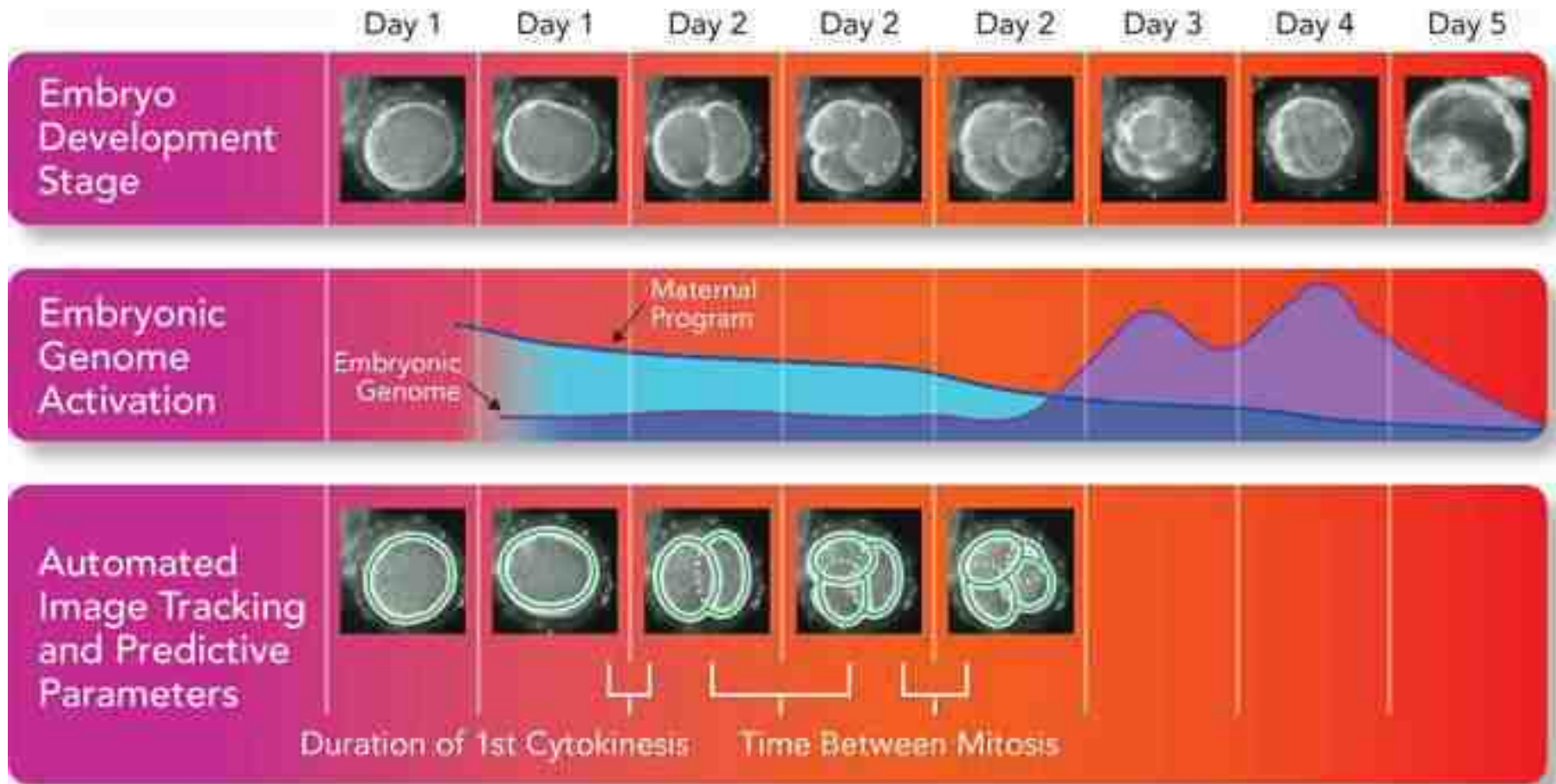
## PRIMO VISION TIME-LAPSE EMBRYO MONITORING SYSTEM





# Auxogyn

- Using a Day 2 algorithm they can predict with over 90% efficiency which day 2 embryo will develop to a blastocyst
- Wong et al. Nat Biotechnol. 2010

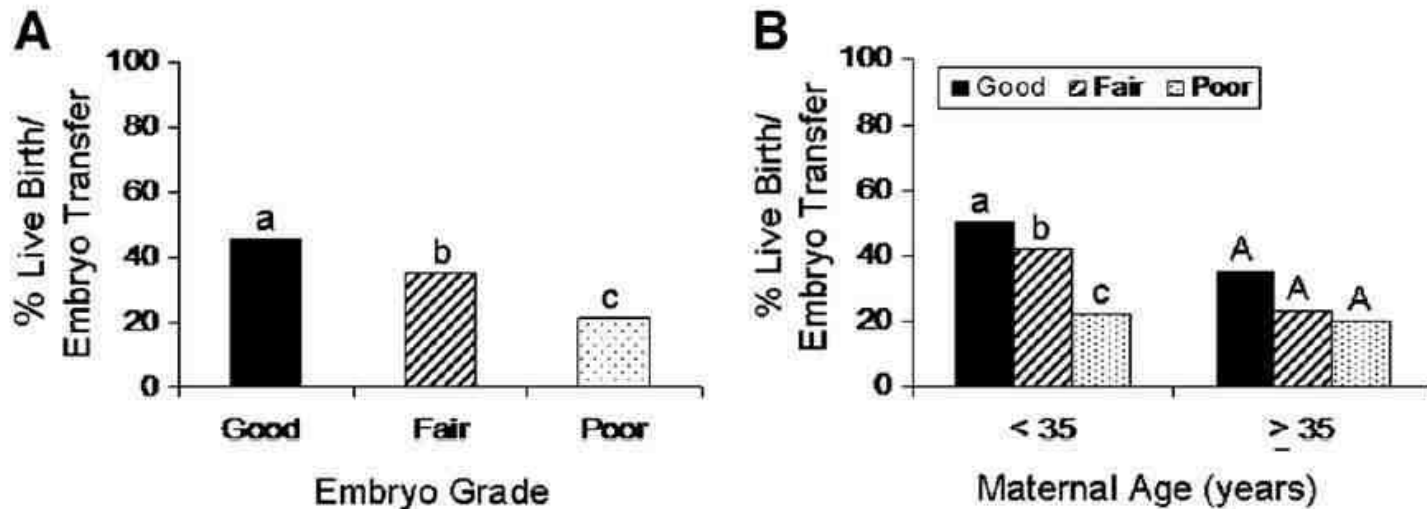


**Is there still a place for  
morphology and will real time  
imaging replace it?**

# Utility of the national embryo morphology data collection by the Society for Assisted Reproductive Technologies (SART): correlation between day-3 morphology grade and live-birth outcome

**FIGURE 1**

(A) Relationship between embryo quality (grade) and the percentage of live births/embryo transfers in patients receiving two day-3 embryos of the same grade. Embryo quality (grade) was positively correlated with the percentage of live births/embryo transfer. Groups with different letters indicate statistical difference ( $P < .0001$ ). (B) Relationship between embryo quality (grade) and maternal age in patients in whom two day-3 embryos of the same grade were transferred. Embryo quality (grade) was positively correlated with the percentage of live births/embryo transfer for the  $< 35$  year olds ( $P = .011$ ), with a positive trend (not statistically significant) with the  $> 35$  year olds ( $P = .299$ ). Groups with different letters are statistically significantly different from one another.



## Trophectoderm morphology: an important parameter for predicting live birth after single blastocyst transfer

A. Ahlström\*, C. Westin, E. Reisner, M. Wikland, and T. Hardarson

Trophectoderm morphology predicts live birth

3293

**Table III** Effect of Expansion, ICM and TE separately analysed and adjusted for known and significant confounders (age, GQE, FSH total dose and number of earlier cycles).

Variable	Value	Live birth, % (n)	OR (95% CI)	P-value
Expansion	0+1+2	23.7 (58)		
	3	36.4 (148)	1.57 (1.07–2.29)	0.0196
	4	46.5 (216)	1.38 (1.04–1.83)	0.0259
inner cell mass	A	45.3 (301)		
	B	31.0 (62)	0.62 (0.44–0.89)	0.0082
	C	14.3 (1)	0.38 (0.05–3.05)	NS
Trophectoderm cells	A	49.9 (234)		
	B	33.9 (128)	0.36 (0.41–0.74)	0.0001
	C	8.0 (2)	0.17 (0.04–0.76)	0.0198

GEE models have been used as they allow for adjustment of within-individual correlations.

For Expansion OR for 3 versus 0+1+2 and 4+5 versus 3 are presented.

For ICM and TE cells OR for 1 versus 0 and 2 versus 3 are presented.

Expansion: 0 was categorized together with 1 due to very few observations with 0 category and no occurrence of live birth in 0 category; 2 was categorized together with 0 and 1 due to no significant difference in the step 0+1 versus 2 in the analysis.

**Table 1 Patient and morphology characteristics of transferred embryos divided by live birth outcome.**

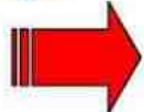
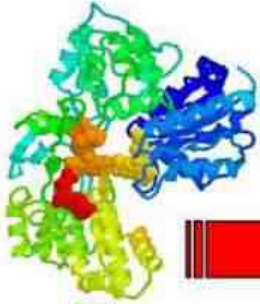
Variable	No live birth (n = 695)	Live birth (n = 422)
Expansion <sup>a</sup> , (n) %		
0	3.0 (21)	0.0 (0)
1	6.6 (46)	5.7 (24)
2	17.3 (120)	8.1 (34)
3	37.3 (259)	35.1 (148)
4	35.8 (249)	51.2 (216)
Inner cell mass <sup>a</sup> , (n) %		
A	71.7 (364)	82.7 (301)
B	27.2 (138)	17.0 (62)
C	1.2 (6)	0.3 (1)
Trophectoderm cells <sup>a</sup> , (n) %		
A	46.3 (235)	64.3 (234)
B	49.2 (250)	35.2 (128)
C	4.5 (23)	0.5 (2)
No score for ICM and TE <sup>a</sup>	187	58

# Morphology and Real Time Imaging

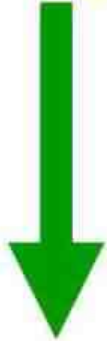
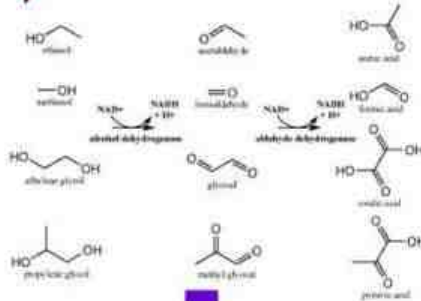
- Morphology has been assisting embryologists for many years to improve embryo selection.
  - It may be logical that by providing more detailed information concerning cleavage patterns and times that morphokinetics has the potential to improve the chance of live birth
  - No large RCT has yet compared MORPHOKINETICS against SET with blastocyst

# Non-Invasive: Part 2

Proteins



Metabolites

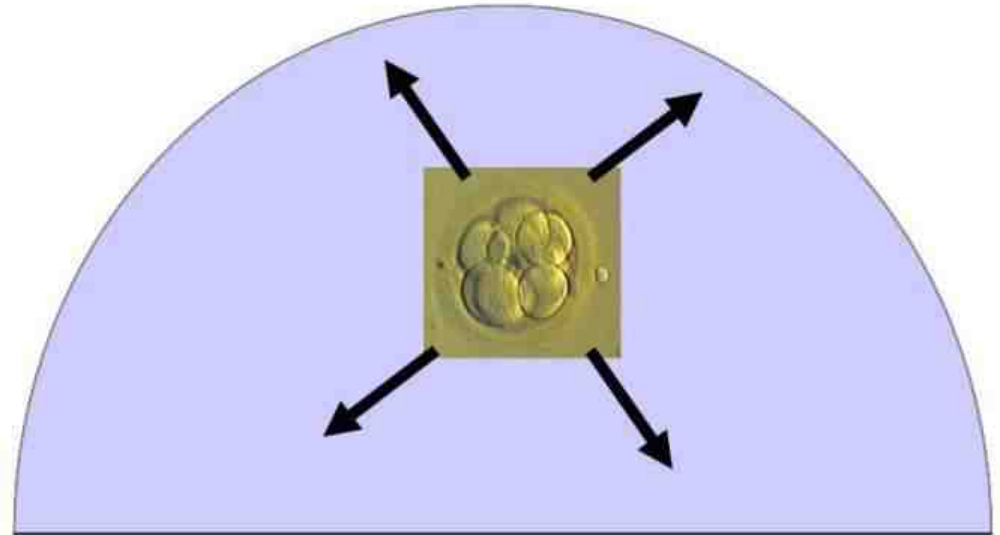


Proteome



Metabolome

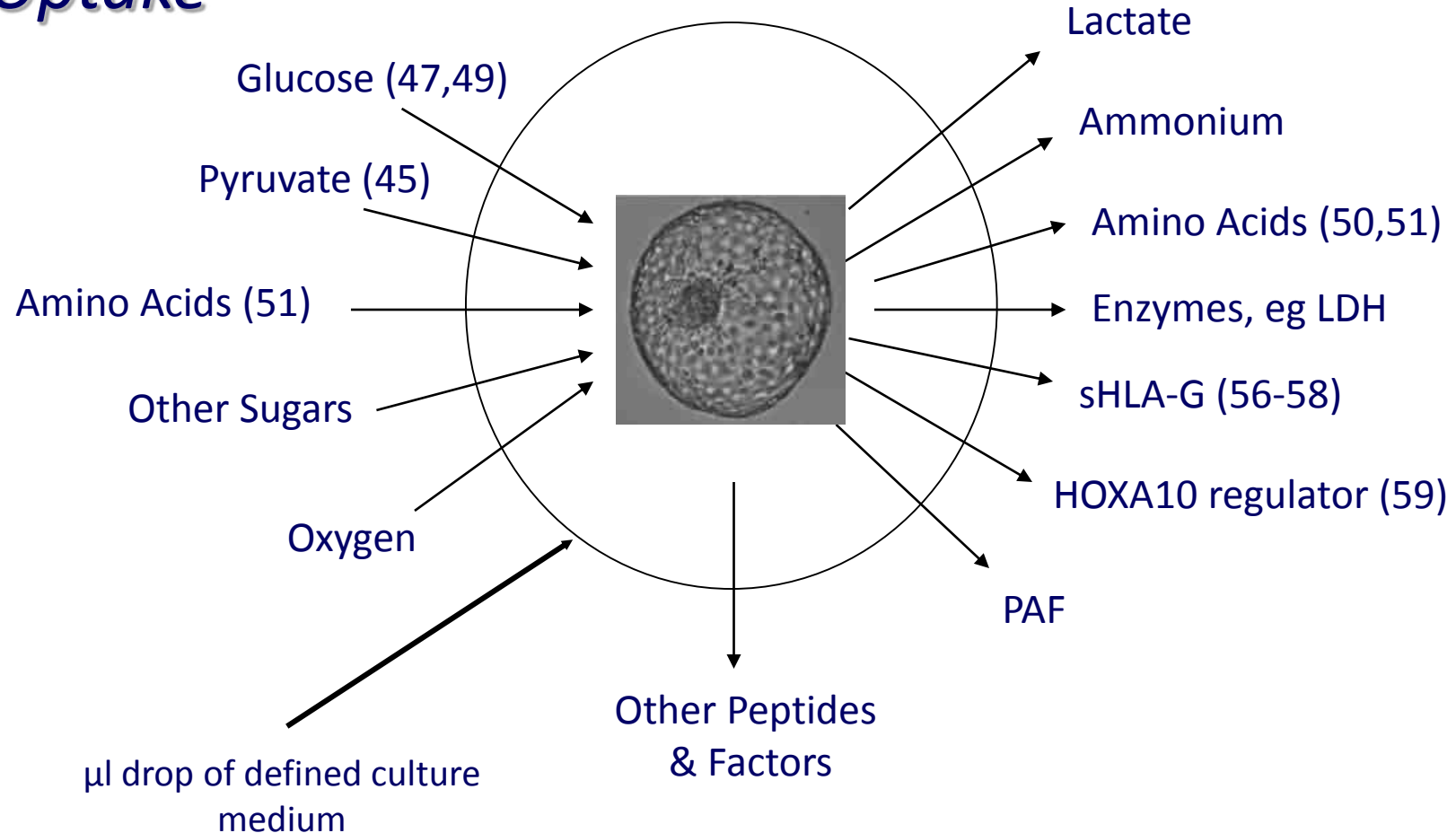
NON-INVASIVE



# CHANGES IN THE CULTURE MEDIA INDICATING THE VIABILITY OF EMBRYOS

*Uptake*

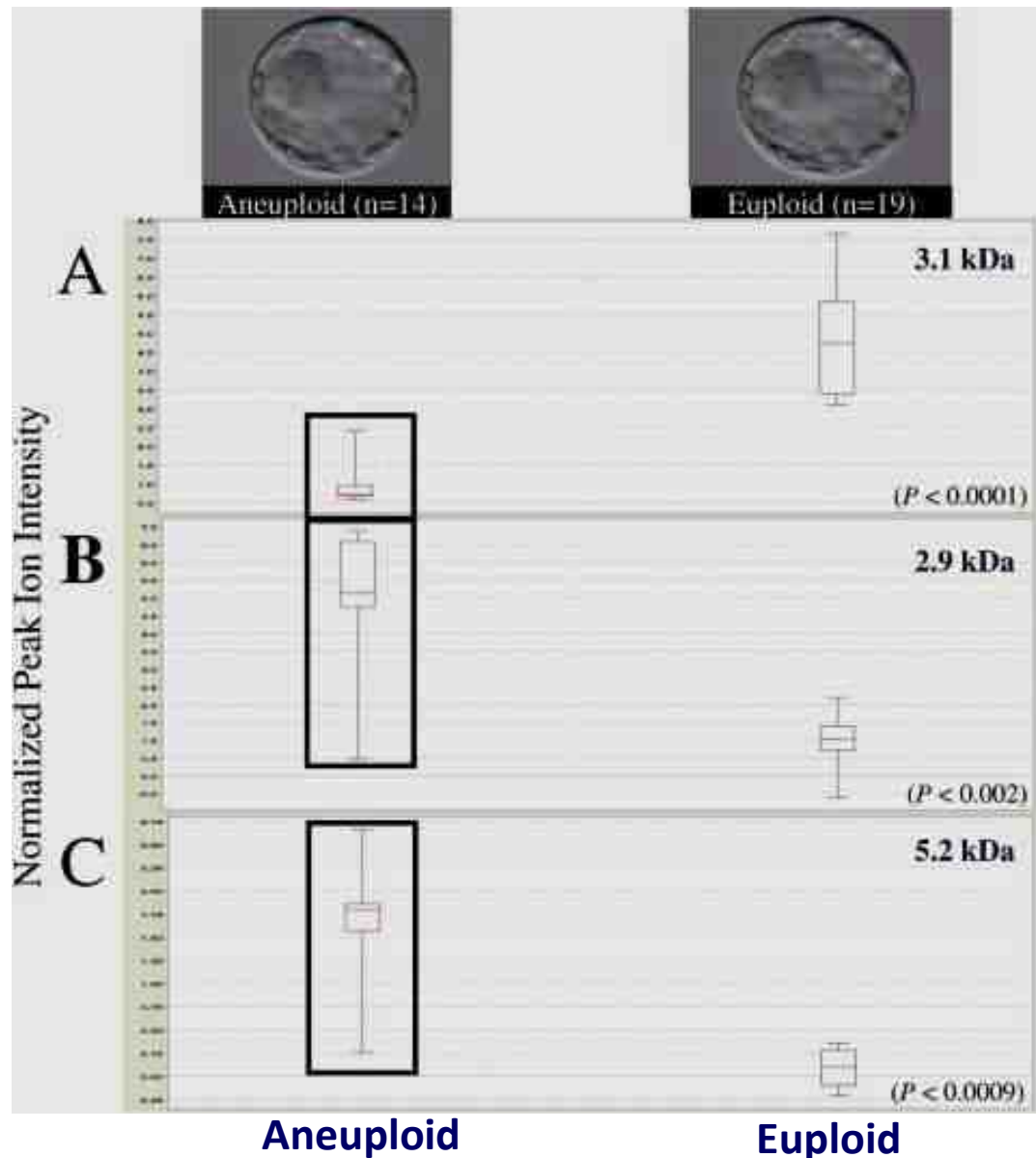
*Production*



[Gardner and Leese, Handbook of IVF. 2000]

[Sakkas and Gardner, Curr. Opin. Obstet. Gynecol. 2005]

# PROTEOMICS AND ANEUPLOIDY



Examples of biomarkers that were differentially expressed in the secretome signatures of euploid blastocysts ( $n = 19$ ) compared with the secretome signature of aneuploid blastocysts ( $n = 14$ ) ( $P < 0.05$ ).

# Other Candidates

## The effect of the biochemical marker soluble human leukocyte antigen G on pregnancy outcome in assisted reproductive technology—a multicenter study

Dirk Kotze, Ph.D.,<sup>a</sup> Thinus F. Kruger, M.D., Ph.D., D.Sc.,<sup>b</sup> Carl Lombard, Ph.D.,<sup>c</sup> Trishanta Padayachee, M.Sc.,<sup>c</sup> Levent Keskinetepe, D.V.M., Ph.D.,<sup>d</sup> and Geoffrey Sher, M.B.Ch.B.<sup>e</sup>

**TABLE 1**

Mean age, mean number of embryos returned, number of chemical pregnancies, clinical pregnancies, and ongoing pregnancies by site and sHLA-G test result for individuals with day-3 transfers only.

Site	sHLA-G	Mean age (y)	Mean no. of embryos <sup>a</sup>	Chemical pregnancy <sup>b</sup>	Clinical implantation <sup>b</sup>	Multiple clinical <sup>b</sup>	Ongoing pregnancy <sup>b</sup>	Multiple ongoing <sup>b</sup>	Total (n)
A	Negative	35.1	1.8	246 (79.7)	222 (71.0)	103 (33.1)	220 (70.7)	101 (32.5)	311
	Positive	34.7	2.0	55 (17.5)	46 (59.0)	22 (28.2)	44 (56.4)	22 (28.7)	78
B	Negative	35.3	1.7	103 (80.8)	178 (75.5)	81 (34.8)	176 (75.5)	79 (33.8)	201
	Positive	34.6	1.7	87 (55.1)	80 (50.6)	47 (29.7)	72 (45.6)	38 (24.1)	158
C	Negative	33.9	2.8	33 (50.8)	31 (47.7)	15 (23.1)	27 (41.5)	9 (13.8)	65
	Positive	35.1	2.6	54 (58.1)	49 (52.7)	22 (24.4)	45 (48.4)	29 (31.7)	93
D	Negative	33.7	3.0	238 (61.5)	225 (58.1)	57 (14.7)	202 (52.5)	44 (11.4)	367
	Positive	33.1	2.1	57 (38.0)	49 (32.7)	14 (9.3)	44 (29.3)	7 (4.7)	150
E	Negative	34.0	2.9	181 (76.4)	176 (74.3)	43 (18.1)	158 (66.7)	37 (15.6)	237
	Positive	36.6	1.7	53 (38.4)	48 (34.0)	21 (15.2)	47 (34.1)	19 (13.8)	138
F	Negative	39.0	2.2	8 (25.5)	8 (23.5)	1 (2.9)	7 (20.6)	0 (0.0)	34
	Positive	35.8	1.6	45 (83.3)	40 (38.5)	20 (19.2)	40 (38.5)	18 (18.3)	104
G	Negative	32.5	3.3	282 (87.9)	248 (42.1)	53 (9.0)	206 (36.0)	26 (4.4)	589
	Positive	38.0	3.0	178 (88.0)	101 (33.3)	13 (4.3)	65 (21.5)	4 (1.3)	303
H	Negative	37.0	2.4	143 (57.0)	147 (51.0)	40 (14.0)	141 (49.3)	22 (7.7)	288
	Positive	35.2	2.6	102 (87.7)	88 (41.7)	31 (14.5)	72 (33.0)	26 (12.7)	214
I	Negative	35.1	2.7	50 (81.8)	23 (38.5)	5 (8.3)	8 (8.5)	3 (3.2)	94
	Positive	35.2	2.5	72 (60.0)	65 (54.2)	28 (23.7)	64 (53.3)	23 (19.2)	126
Total		35.7	2.9	1,010 (56.2)	811 (50.7)	312 (17.4)	818 (45.0)	254 (14.7)	1,797

<sup>a</sup> Mean number of embryos transferred.

<sup>b</sup> Count percentage of total.

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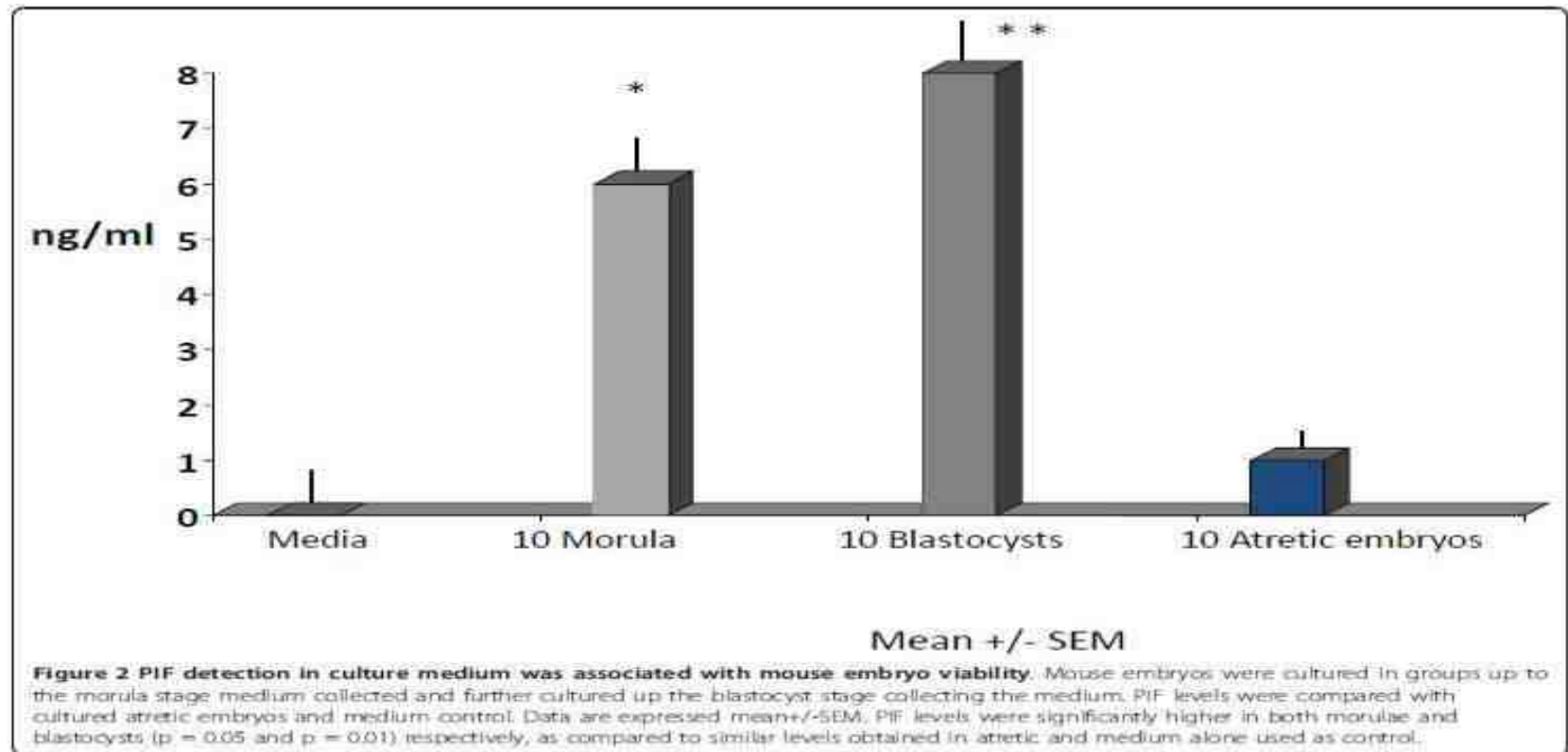
# Other Candidates

## RESEARCH

## Open Access

### Preimplantation Factor (PIF) correlates with early mammalian embryo development-bovine and murine models

Christopher W. Stamatkin<sup>1</sup>, Roumen G. Roussev<sup>1</sup>, Mike Stout<sup>2</sup>, Victor Absalon-Medina<sup>3</sup>, Sivakumar Ramu<sup>1</sup>, Chelsi Goodman<sup>1</sup>, Carolyn B. Coulam<sup>1</sup>, Robert O. Gilbert<sup>3</sup>, Robert A. Godke<sup>2</sup> and Eytan R. Barnea<sup>4,5\*</sup>

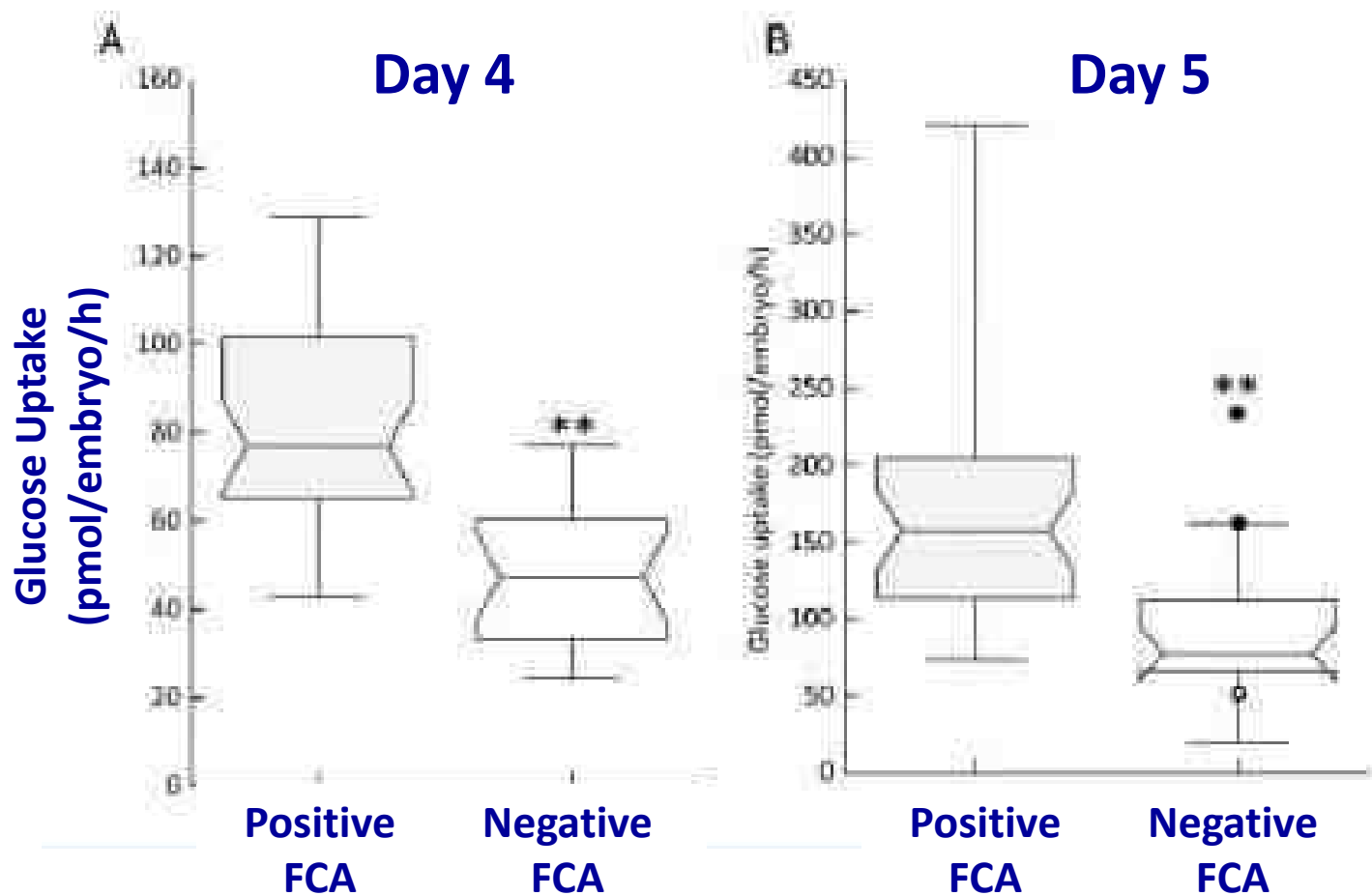


# The Metabolism of Embryos

- **Assessment of specific metabolic pathways, their relative activities and their regulation in relation to embryo viability**
- **Example:**
  - **Glucose, Lactate, Pyruvate**
  - **TCA Cycle functions before 8-cell stage and**
  - **Glycolysis functions after the 8-cell stage**

**It was first shown in 1980 that Glucose metabolism is linked with the viability of embryos**

**Glucose consumption of single post-compactation human embryos is predictive of embryo sex and live birth outcome. (Gardner et al., Human Reproduction 2012)**



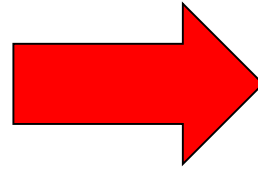
# Clinical Performance Summary of Metabolomics by Near Infra Red

TYPE OF NIR INSTRUMENT	STUDY TYPE		MORPHOLOGY	MORPHOLOGY PLUS VIAMETRICS (NIR)	BENEFIT
<b>Prototype</b> <i>Hardarson et al.</i> <i>(Human Reprod, 2012)</i>	<b>Single Embryo Transfer</b>	Live Birth Rate	Day 2: 22/83 (26.5%) Day 5: 36/80 (45.0%)	Day 2: 27/ 87 (31.0%) Day 5: 30/77 (39.0%)	YES  NO
<b>Prototype</b> <i>Vergouw et al.</i> <i>(Human Reprod, 2012, 2014)</i>	<b>Single Embryo Transfer</b>	Live Birth Rate	Day 3: 68/163 (41.7%)	Day 3: 61/146 (41.8%)	NO
<b>Commercial</b> <i>Economou et al.</i> <i>(ESHRE)</i>	<b>Double Embryo Transfer</b>	Clinical Pregnancy Rate	8/28 (29% )	16/28 (57%)	YES
<b>Commercial</b> <i>Sfontouris et al.</i> <i>(J Reprod Fertil 2013)</i>	<b>Multiple Embryo Transfer</b>	Clinical Pregnancy and Implantation Rate	41/86 (47.7%) 66/257 (25.7%)	21/39 (53.9%) 35/102 (34.3%)	YES

# The Non- invasive Proteomics/ Metabolomics

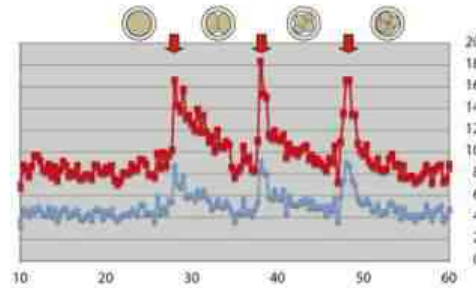
## Example:

### A Strong Hypothesis Can Be A Slave To Technology



INCONSISTENT  
RESULTS

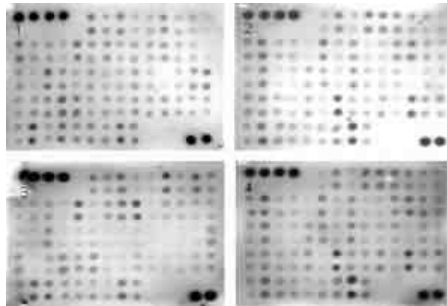
# EMBRYO ASSESSMENT FOR SINGLE EMBRYO TRANSFER



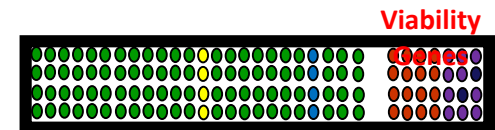
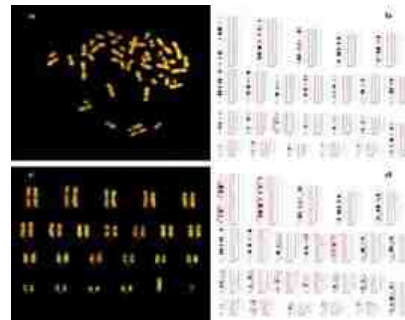
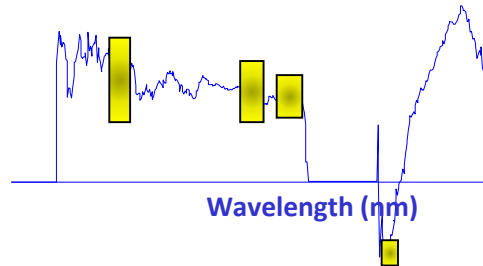
Real Time  
Morphology



Metabolic  
assessment  
or imaging



Proteomics



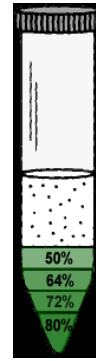
Viability

Single  
Gene  
Defects

Transcriptomics and Genomics

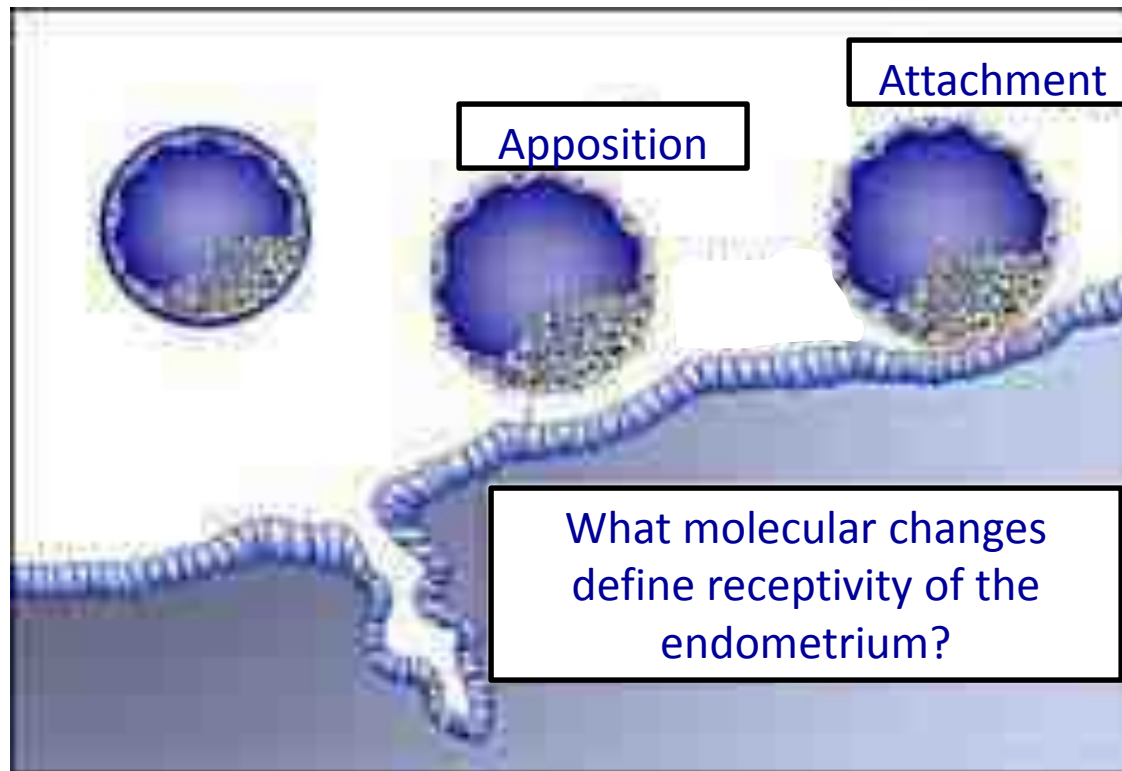
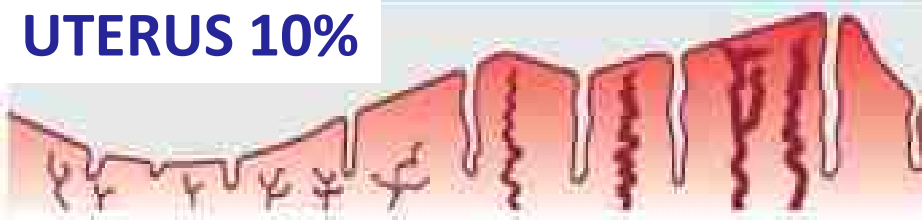
# Selection Methods to isolate the best sperm Proteomics

- Density – *Gradient Separation*
- Surface Charge – *Electrophoresis, Zeta Method*
- Morphological Characteristics – *IMSI*
- Motility Characteristics – *Zech Selector, Microfluidics*
- Membrane Integrity – *Hyaluronan Binding, HOST*
- Surgical – *Testicular Surgery*



# Venn diagram of the responsibilities of Reproduction Failure:

UTERUS 10%

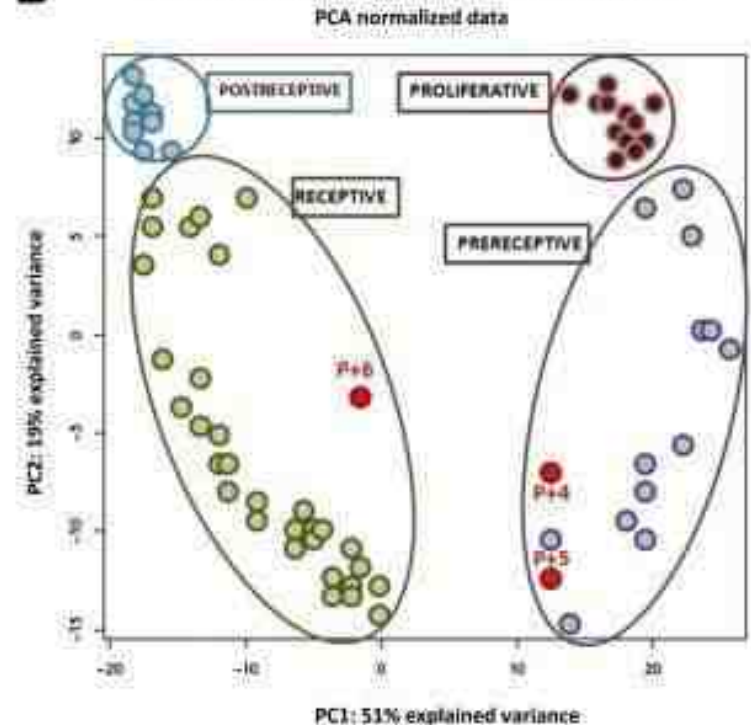


# The endometrial receptivity array for diagnosis and personalized embryo transfer as a treatment for patients with repeated implantation failure

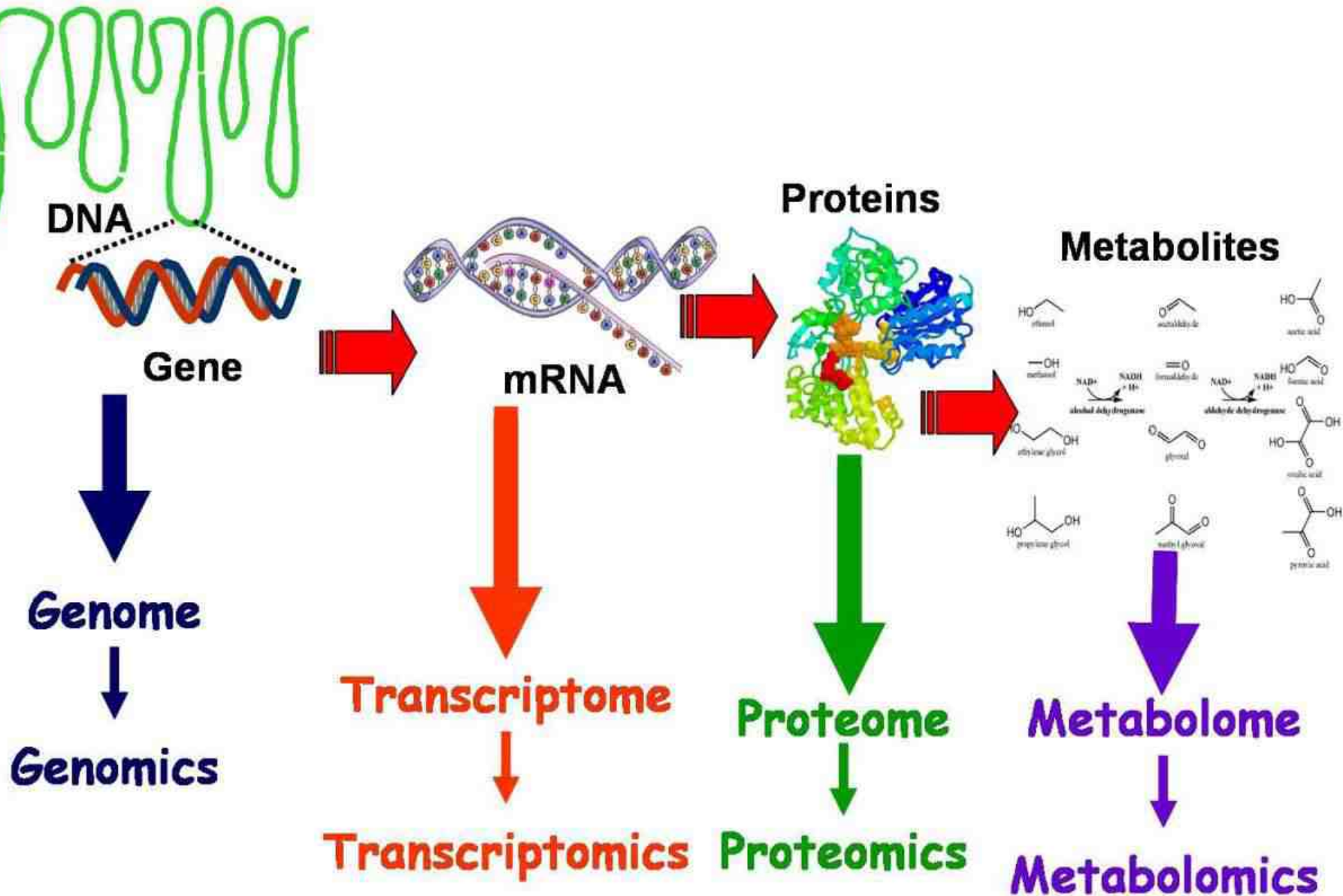
**A** Clinical outcome of non receptive RIF and control patients that underwent pET

	Non Receptive
No. of patients	25
No. of previous failed cycle RIF Patients	5.0±1.8
No. of previous failed cycle Control Patients	0.3±0.6
ERA Prediction	
Pre-receptive	21/25 (84.0)
Post-receptive	4/25 (16.0)
2 <sup>nd</sup> ERA at the specified day (P+4;P+6;P+7;LH+9) <sup>a</sup>	18
Months between 1 <sup>st</sup> and 2 <sup>nd</sup> ERA	2.6±2.8
2 <sup>nd</sup> ERA Receptive at the specified day	15
Patients with pET <sup>b</sup> after 2 <sup>nd</sup> RECEPTIVE ERA	8
Months between 2 <sup>nd</sup> RECEPTIVE ERA and pET	1.8±0.7
Implantation rate using pET	5/13 (38.5)
Pregnancy rate using pET	4/8 (50.0)
Biochemical pregnancies (%)	0/4 (0.0)
Clinical abortions (%)	0/4 (0.0)

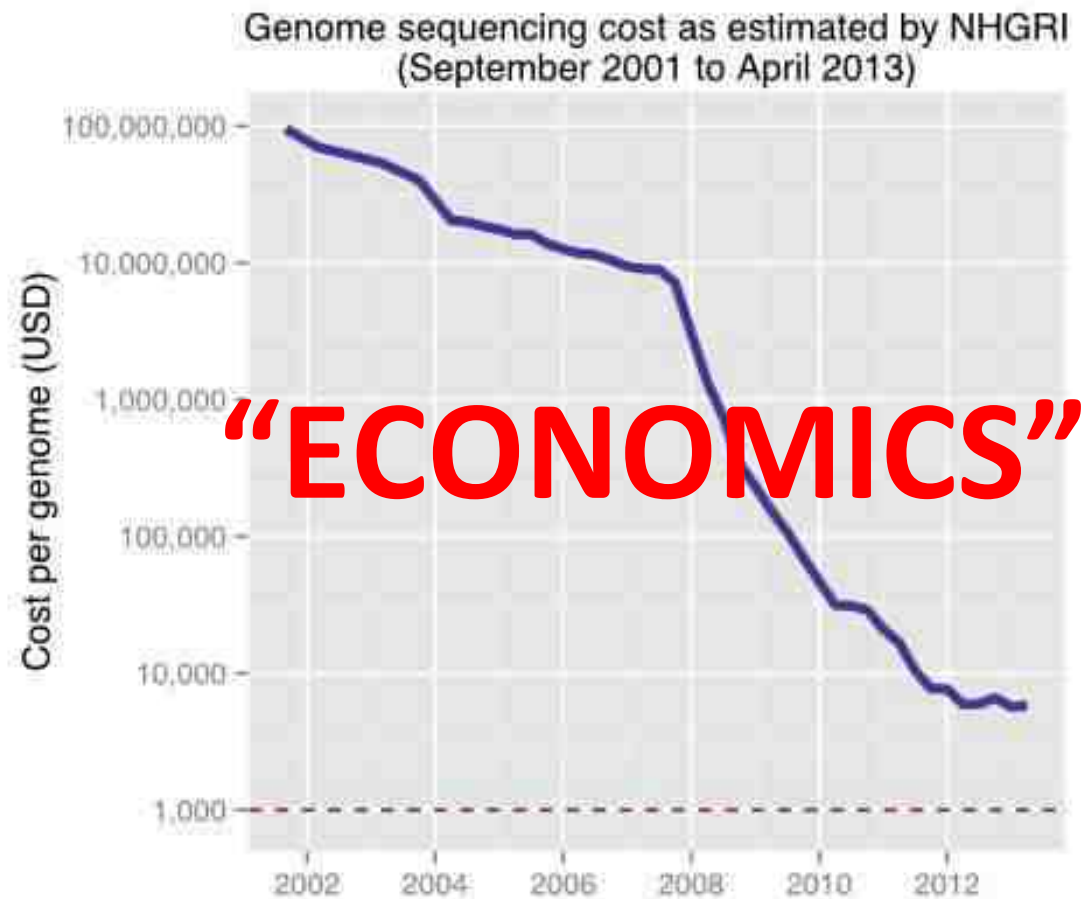
**B** Principal component analysis with ERA samples



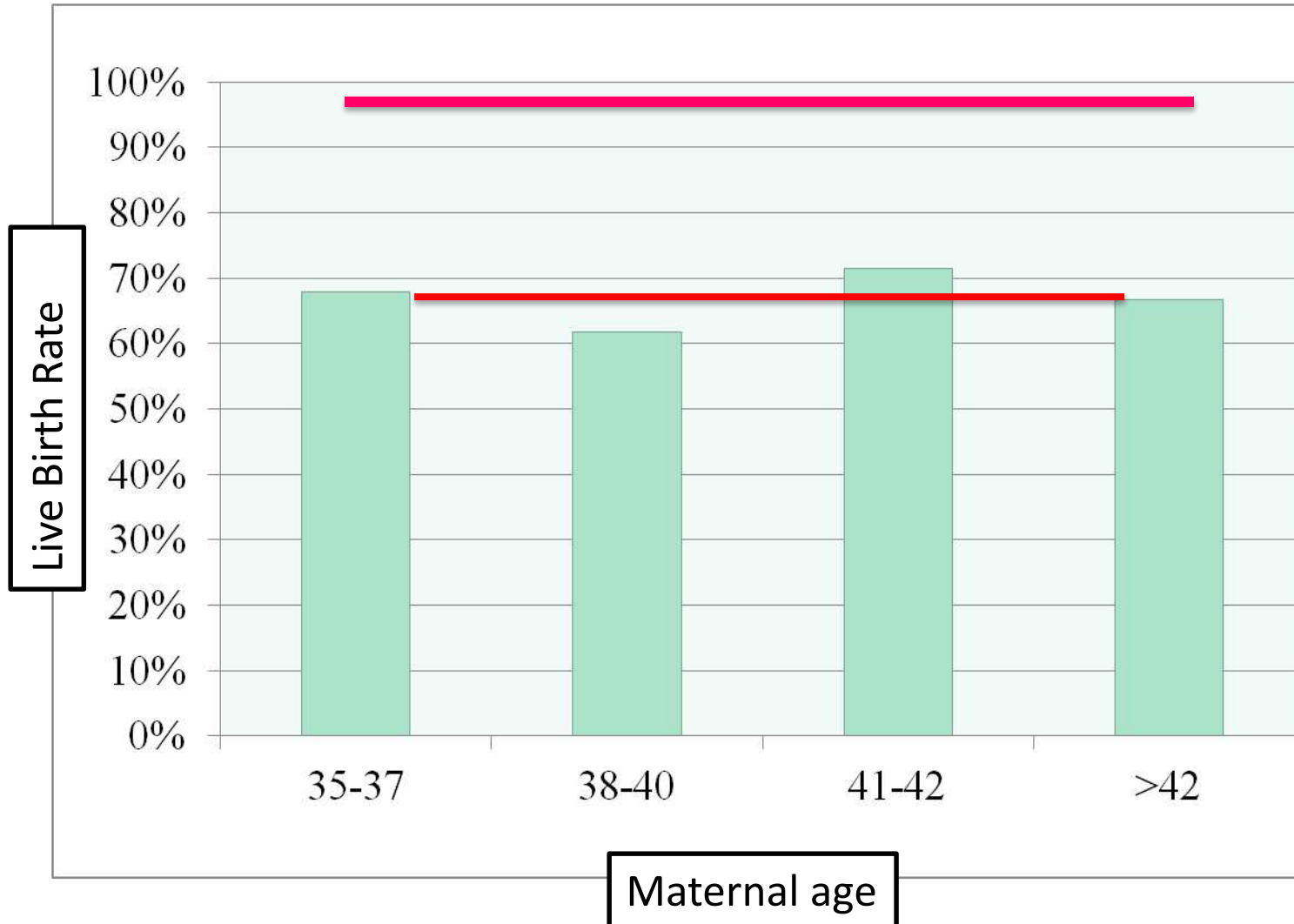
# The “Omes and Omics”



# THE FINAL AND THE BIGGEST “OMICS”



# OMICS technologies used to improve Singleton Live Birth Rates



**Thank you**