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First for Biological Science in UK

MSc Human Clinical Embryology and Assisted Conception

The MSc in Human Clinical Embryology and Assisted Conception is a full-time one-year degree programme focused on the practical and theoretical skills in human embryology and clinical IVF.



Screening of man prior to IVF/ICSI

Declaration of COI

- ❑ Fully employed by University of Dundee (UoD).
- ❑ WHO paid honorarium to be Chair of group and paid travel/accommodation/expenses.
- ❑ Editor in Chief of MHR
- ❑ Grant funding from MRC.
 - UoD Patent – sperm stimulation.
- ❑ Give occasional lectures that are company/society sponsored e.g. Vertex : pay travel/accommodation/expenses.
- ❑ Cambridge University Press – 2 edited books.
- ❑ I'm not on any company board, advisory board or have a single share in anything or anybody.

New – what's history ?

- ❑ Global infertility guidelines:
Strategy for development and dissemination; Generating practice guidelines
- ❑ The revision and updating of the “WHO global Guidelines for infertility diagnosis, management and interventions for treatment” (1992) and the WHO manual for the investigation and diagnosis of the infertile couple” (1993)
- ❑ Initiated in January 2012 committee meeting



Essentials and discussion points

- ❑ Physical examination and history
- ❑ **High quality Semen Analyses**
- ❑ **If abnormal spermatogenesis : potential screens.**

As referral depends on Semen Analysis – make sure its high quality

- Cornerstone - Simple

Potential groups [not a new discovery]

Fertile, Indeterminate and sub fertile ranges and corresponding odds ratio for infertility
 All 3 abnormal. = 15.8 (8.7-29)

Variable	Concentration x10 ⁶ /ml	Motility %	Morphology %
Fertile	>48	>63	>12
Indeterminate	13.5 - 48.0	32 - 63	9 -12
Sub fertile	<13.5 2.2	<32 2.5	<9 2.9

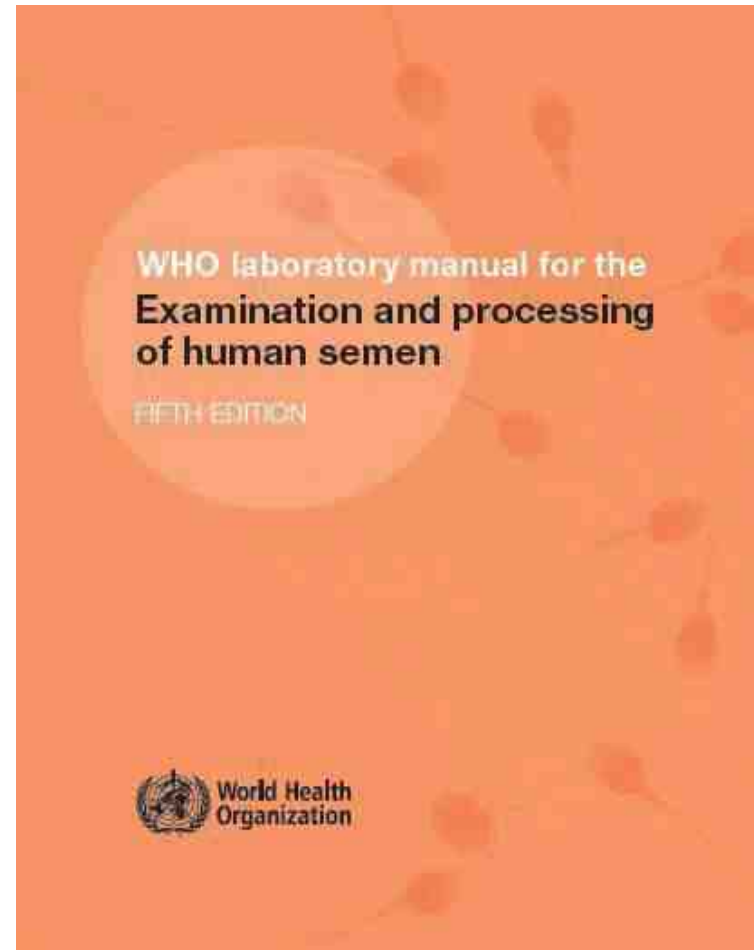
- 696 fertile couples, 765 infertile couples
- Considerable overlap between the groups
- ‘none of the measures are diagnostic of infertility’

•*Minimal values similar to MacLeod and Gold in 1951* “The greatest difference between the two groups [infertile and fertile] is seen at the count levels under 20million/cc. Only 5 per cent of fertile men compared with 16 per cent of the ‘infertile’ group fall into this category” (MacLeod and Gold, 1951). *Almost 60 years ago.....’*

•*Remarkably : Data similar to new WHO 2009*
 Guzick et al., (2001) NEJM 345, 1388-1399

WHO Semen analysis

- ❑ Volume
- ❑ Concentration
- ❑ Motility
- ❑ Morphology



Can we count sperm?

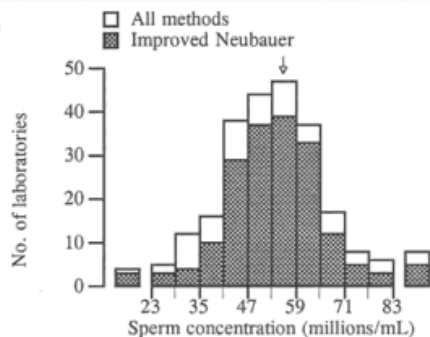
The quality of the assessment is almost always poor....

Semen analysis has a high degree of error

[or more correctly the person doing it does].

Specimen S157

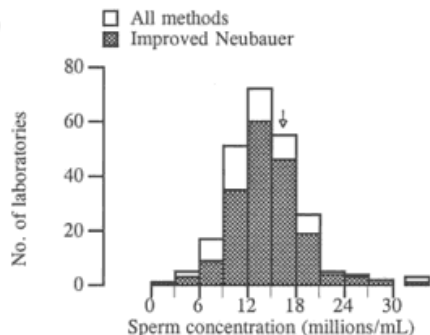
	n	Mean	SD	CV(%)
All methods	242	53.47	9.74	18.2
Horwell	11	47.60	20.34	42.7
Makler	34	49.13	12.30	25.0
Microcell	6	53.34	7.12	13.3
Improved Neubauer	183	54.12	8.82	16.3
Fast Read	1			
Fuchs Rosenthal	2			
Kova	2			
Mika	1			
No counting chamber	1			
Thoma	1			



Your result	57
Designated value	53.47
Your bias (%)	6.60
Your BIS	33
Pool	140

Specimen S158

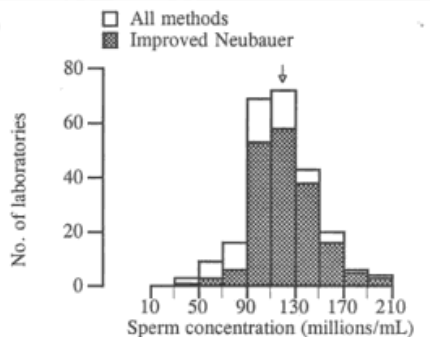
	n	Mean	SD	CV(%)
All methods	241	14.24	3.57	25.1
Horwell	11	13.56	5.03	37.1
Makler	34	13.57	3.14	23.2
Microcell	6	11.90	4.70	39.5
Improved Neubauer	183	14.60	3.24	22.2
Fast Read	1			
Fuchs Rosenthal	2			
Kova	2			
No counting chamber	1			
Thoma	1			



Your result	17
Designated value	14.24
Your bias (%)	19.37
Your BIS	39
Pool	141

Specimen S159

	n	Mean	SD	CV(%)
All methods	242	118.65	19.17	16.2
Horwell	11	95.20	38.22	40.1
Makler	34	106.04	20.31	19.2
Microcell	6	82.50	17.90	21.7
Improved Neubauer	183	121.52	18.02	14.8
Fast Read	1			
Fuchs Rosenthal	2			
Kova	2			
Mika	1			
No counting chamber	1			
Thoma	1			



Your result	117
Designated value	118.65
Your bias (%)	-1.39
Your BIS	-7
Pool	142

Adherence to WHO methods is very poor -
throughout the world.

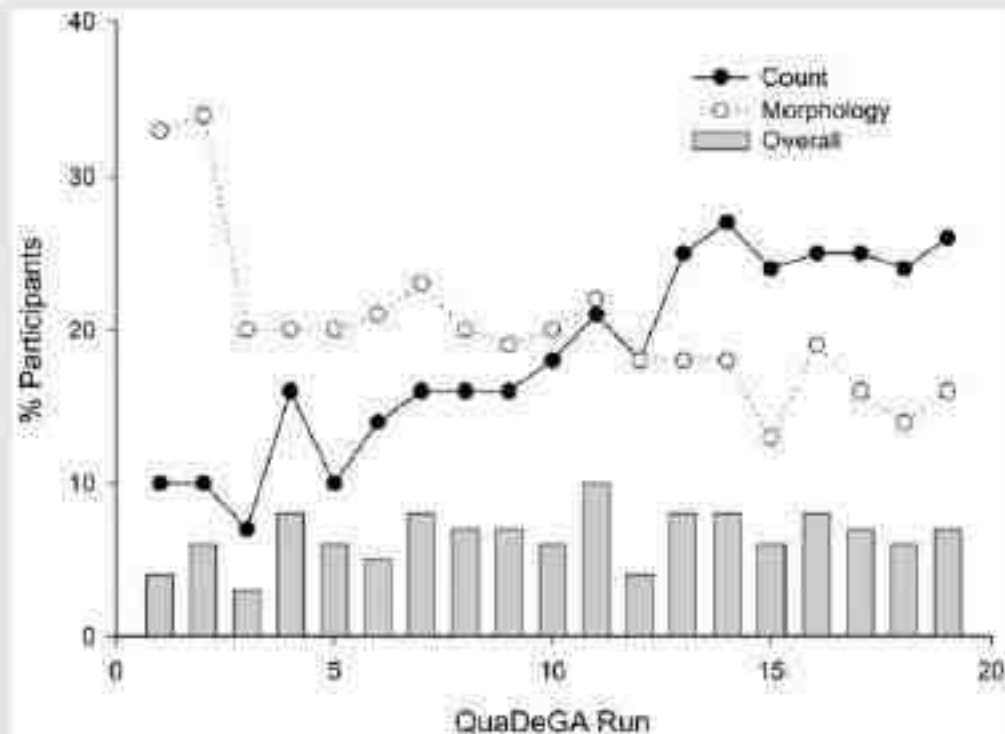
Less than 10% UK laboratories are WHO compliant
 (From Riddell Hum Reprod [1995] 12, 3441-3445)

Table III. Summary of the number of 'specialist laboratories' (SL group) and 'DGH laboratories' (DGH group) that comply with the guidelines suggested in the WHO (1999) manual with regard to morphology assessment

Criteria	SL group (<i>n</i> = 19)	DGH group (<i>n</i> = 18)	Total (<i>n</i> = 37)	Cumulative compliance ^a
Staining methods	7	18	25	25
Papanicolaou or Diff Quick	5	12	17	17
Sperm dimensions	16	16	32	14
×100 oil immersion	3	2	5	4
Count ≥200 sperm	6	5	11	3
External quality control	12	16	28	3
Internal quality control	8	8	16	2
No. of compliant laboratories	1	1	2	2

Similar in Germany...and not improving ...

SUPPLEMENTAL FIGURE 1

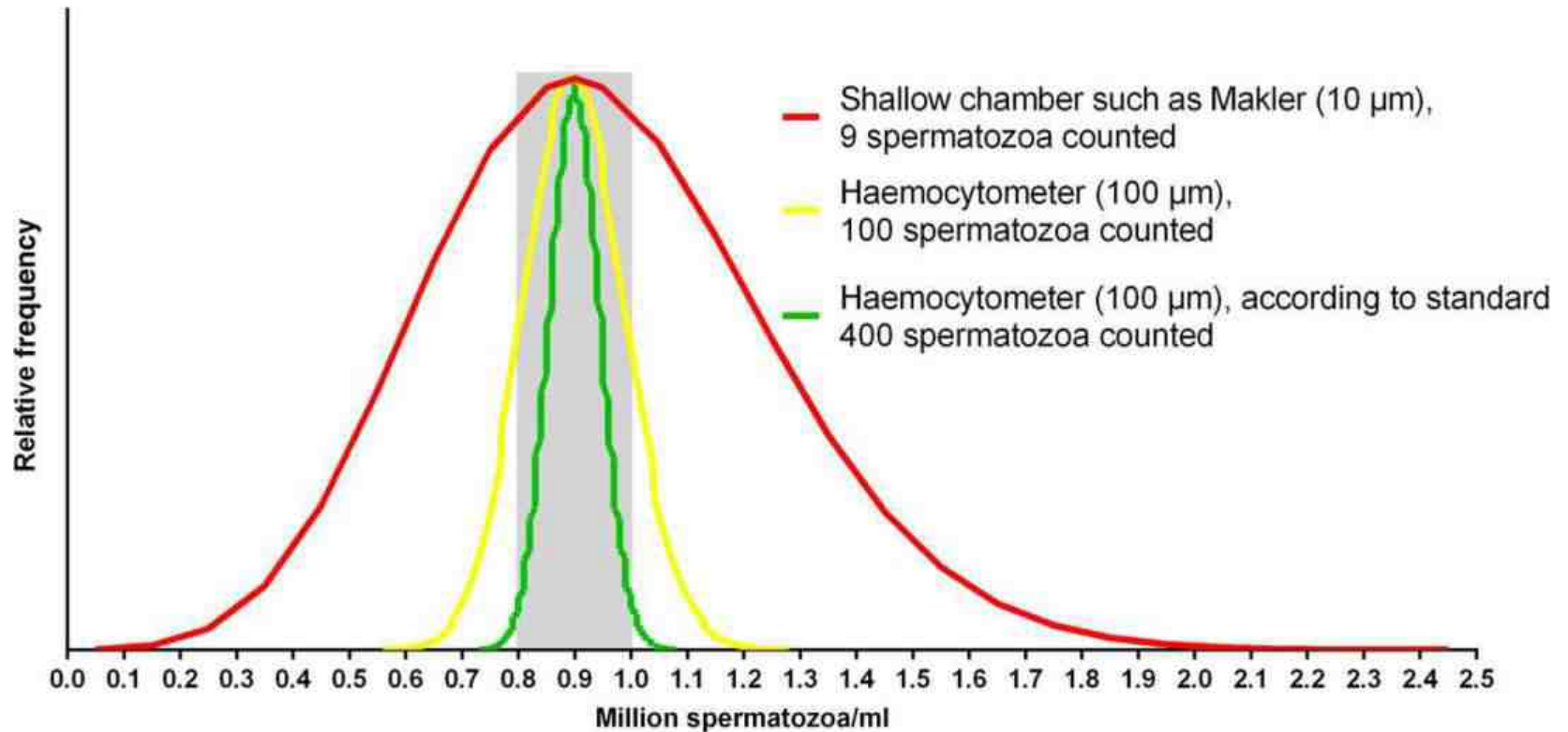


Adherence to WHO guidelines by the QuaDeGA participants in sperm counting (filled circles) and morphology (open circles). The bar graph indicates the percentage of laboratories that follow all the recommended procedures for the evaluation of sperm concentration/number, morphology and motility.

Mallidis. Ten years of semen analysis EQC. Fertil Steril. 2012.

Its important in context of IVF ICSI

A schematic representation of the uncertainties associated with different numbers of spermatozoa counted with results of sperm counting in the borderline zone (grey box)—50:50 IVF/ICSI.



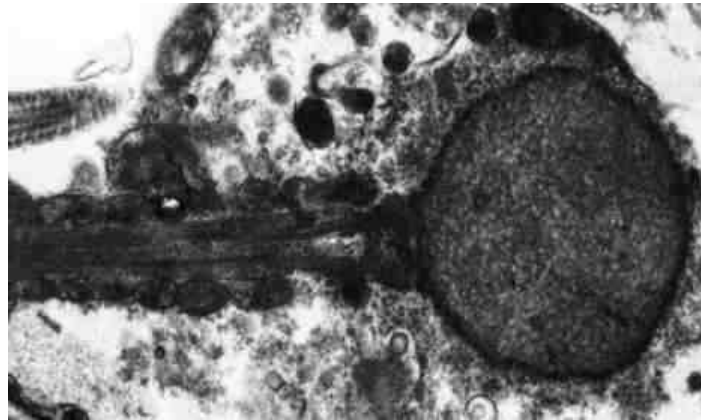
Lars Björndahl et al. *Hum. Reprod.* 2016;31:227-232

human
reproduction

Unfashionable butHow can we improve it?

Perform it with more rigour.....

Not just an issue of numbers - Globozoospermia



- Incidence of globozoospermia 1:10000000 sub-fertile men.
- Generally limited data as few cases reports

Screening - When spermatogenesis goes wrong

- OA vs NOA

NOA not as successful as OA

From Osmanagaoglu *et al.*, Hum Reprod 18, 1836-1840.

K.Osmanagaoglu *et al.*

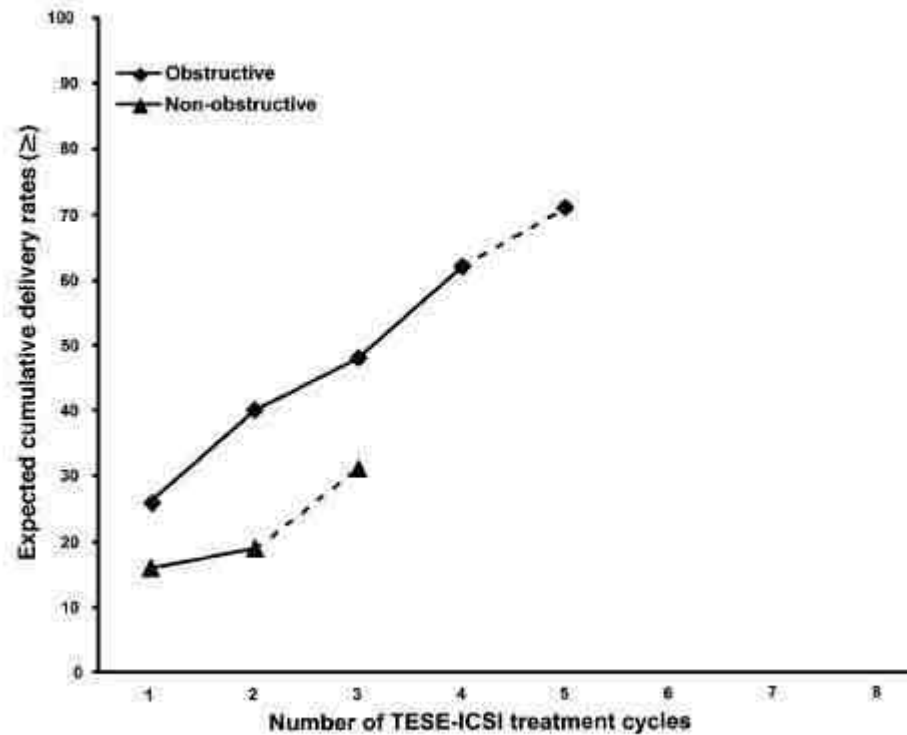


Figure 1. Comparing expected cumulative delivery rates in patients

Table I. Causes of obstruction and spermatogenetic defects

	Obstructive azoospermia [n (%)]	Nonobstructive azoospermia [n (%)]	Hyposperm atogenesis [n (%)]
Anejaculation or retrograde ejaculation	1 (0.3)	1 (0.2)	1 (1.8)
Testicular tumour (benign or malign)	1 (0.3)	3 (0.5)	0 (0)
Inguinal hernia	3 (0.8)	3 (0.5)	0 (0)
Mumps orchitis	0 (0)	9 (1.4)	0 (0)
Congenital bilateral absence of the vas deferens	80 (20.9)	1 (0.2)	4 (7.0)
Post-chemotherapy or radiotherapy	4 (1.0)	24 (3.8)	1 (1.8)
Cryptorchidism	20 (5.2)	99 (15.8)	10 (17.5)
Hydrocele	3 (0.8)	1 (0.2)	0 (0)
Hypogonadotropic hypogonadism	0 (0)	4 (0.6)	0 (0)
Infection	32 (8.4)	19 (3.0)	5 (8.8)
→ Karyotype anomaly	1 (0.3)	65 (10.3)	0 (0)
Failed reversal	88 (23.1)	10 (1.6)	4 (7.0)
Trauma or testicular torsion	3 (0.8)	1 (0.2)	0 (0)
→ Varicocele	9 (2.4)	45 (7.2)	4 (7.0)
Yq deletion	0 (0)	26 (4.1)	0 (0)
Steinert disease	0 (0)	2 (0.3)	2 (3.5)
Congenital anomaly genital tract	3 (0.8)	0 (0)	0 (0)
Diabetic neuropathy	1 (0.3)	0 (0)	0 (0)
Post-vasectomy	12 (3.1)	0 (0)	0 (0)
Toxic	0 (0)	0 (0)	1 (1.8)
Young syndrome	1 (0.3)	0 (0)	0 (0)
Unexplained	119 (31.2)	375 (50.1)	25 (43.8)

From Vermaeue et al., 2006
Hum Reprod 21, 1551-1554

Specific conditions

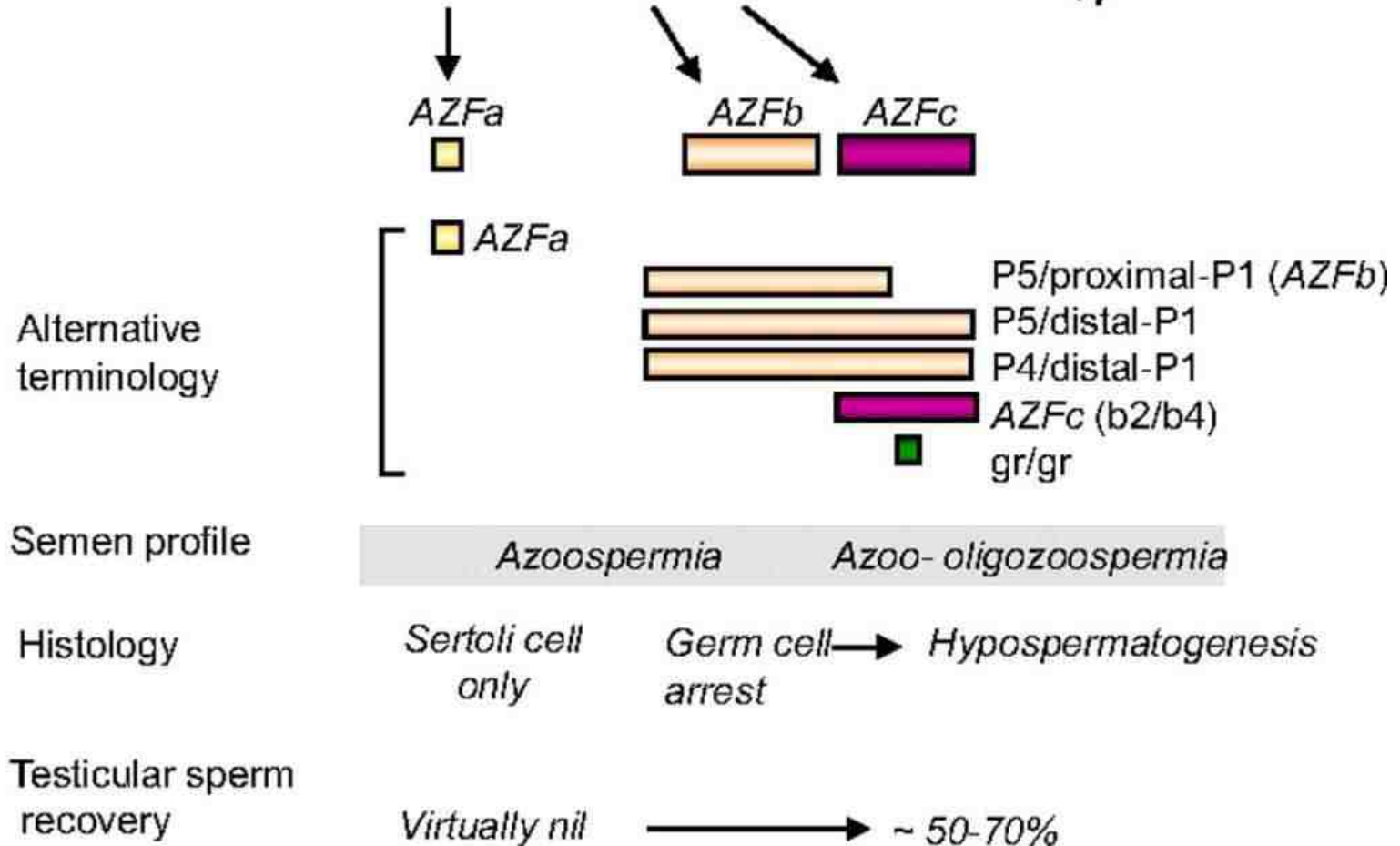
Screening and ART success in Non Mosaic KS

1. Causes still remain to be finally resolved e.g. why reduction in germ cell development?
2. Consistent pregnancies in couples where man has non mosaic KS using testicular cells. At least 101 children born (Fullerton)
3. Prediction of recovery and success remains difficult. Recovery ~44% of the time. Most predictive factor - age of man at recovery (younger being better).
4. Suggestion 3 groups of patients :
 1. With focal spermatogenesis
 2. No sperm but spermatogonia
 3. No germ cells at all
5. The majority of births healthy.



Possible use of Y deletions....

Yq microdeletions.



McLachlan R I , O'Bryan M K JCEM 2010;95:1013-1024

Y deletions provide valuable information recovery of sperm

TABLE 2				
Outcomes of microdissection TESE in azoospermic men stratified by Y microdeletion status.				
Etiology of azoospermia	Sperm retrieved	Sperm not retrieved	Total	Retrieval rate
AZFa	0	2	2	0%
AZFb	0	7	7	0%
AZFb+c	0	7	7	0%
AZFa+b+c	0	4	4	0%
AZFc	15	6	21	71.4% ^a
Nondeleted, idiopathic	188	197	385	48.8% ^a

Note: TESE = testicular sperm extraction; AZF = azoospermic factor.
^a Comparison of retrieval rates in AZFc deleted men and idiopathically azoospermic nondeleted men, $P < .05$ (Fisher's exact test).

Stahl. Y microdeletion screening is essential. Fertil Steril 2010.

Y deletions provide valuable information – success

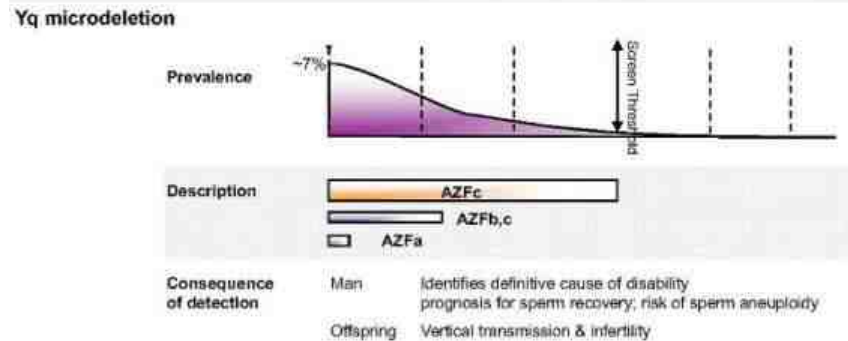
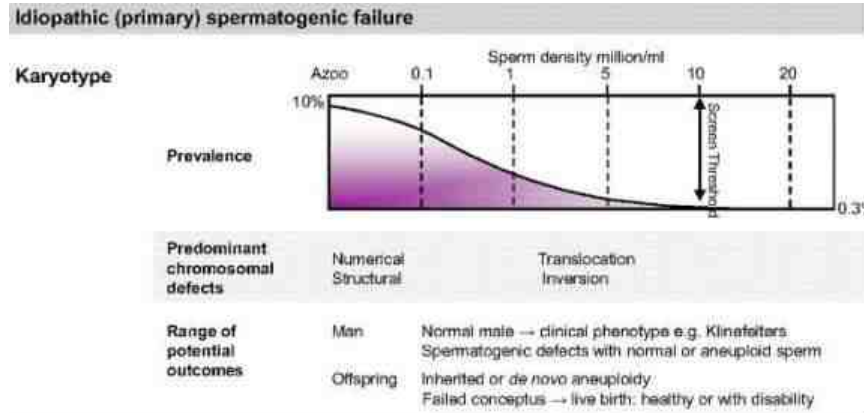
TABLE 3			
Clinical pregnancy outcomes in idiopathically azoospermic men and in azoospermic AZFc deleted men in whom sperm were surgically retrieved.			
Etiology of azoospermia	Sperm retrieved	Clinical pregnancies	Clinical pregnancy rate
Idiopathic	188	91	48.4% ^a
AZFc deletion	15	10	66.7% ^a

^a Not significantly different ($P = .19$, Fisher's exact test).

Stahl. Y microdeletion screening is essential. Fertil Steril 2010.

Potential summary

Essential genetic investigations in male infertility – a Model – 2010.



Obstructive azoospermia

CFTR mutation	Cystic Fibrosis	BCAV / idiopathic epididymal
Genotype	homozygous	heterozygote / compound heterozygote
Mutation	Severe coding	Milder coding / 5T allele
Consequence	Clinical CF	Renal anomalies / sinopulmonary disease
		Female partner screen Residual CF risk / PGD

McLachlan R I , O'Bryan M K JCEM 2010;95:1013-1024

Whilst data limited there appears no significant cause for concern over health of the children

Neonatal outcome

Belva *et al.*, (2011) *Hum Reprod* 26, 1752-1758

Table III Low birthweight, very low birthweight and prematurity rates according to sperm origin.

	Testicular sperm ^a (n = 512)	Epididymal sperm ^a (n = 182)	Ejaculated sperm (n = 2477)
Low birthweight			
Singletons	26/314 (8.2)	5/96 (5.2)	101/1299 (7.8)
Multiples			
Twin	92/168 (55.4)	32/55 (58.1)	588/1098 (53.5)
Triplet	16/21 (76.2)	17/18 (94.4)	70/80 (87.5)
Very low birthweight			
Singletons	4/314 (1.3)	0/96 (0)	21/1299 (1.6)
Multiples			
Twin	8/168 (4.8)	4/55 (7.3)	54/1098 (4.9)
Triplet	5/21 (23.8)	4/18 (22.2)	30/80 (37.5)
Prematurity			
Singletons	32/316 (10.1)	12/93 (12.9)	126/1299 (9.7)
Multiples			
Twin	104/160 (65)	33/58 (56.9)	647/1098 (58.9)
Triplet	21/21 (100)	21/21 (100)	80/80 (100)

Data are given as numbers (%).

^aNo difference was found comparing low birthweight, very low birthweight and prematurity rate in the testicular and epididymal sperm group versus the ejaculated sperm group.

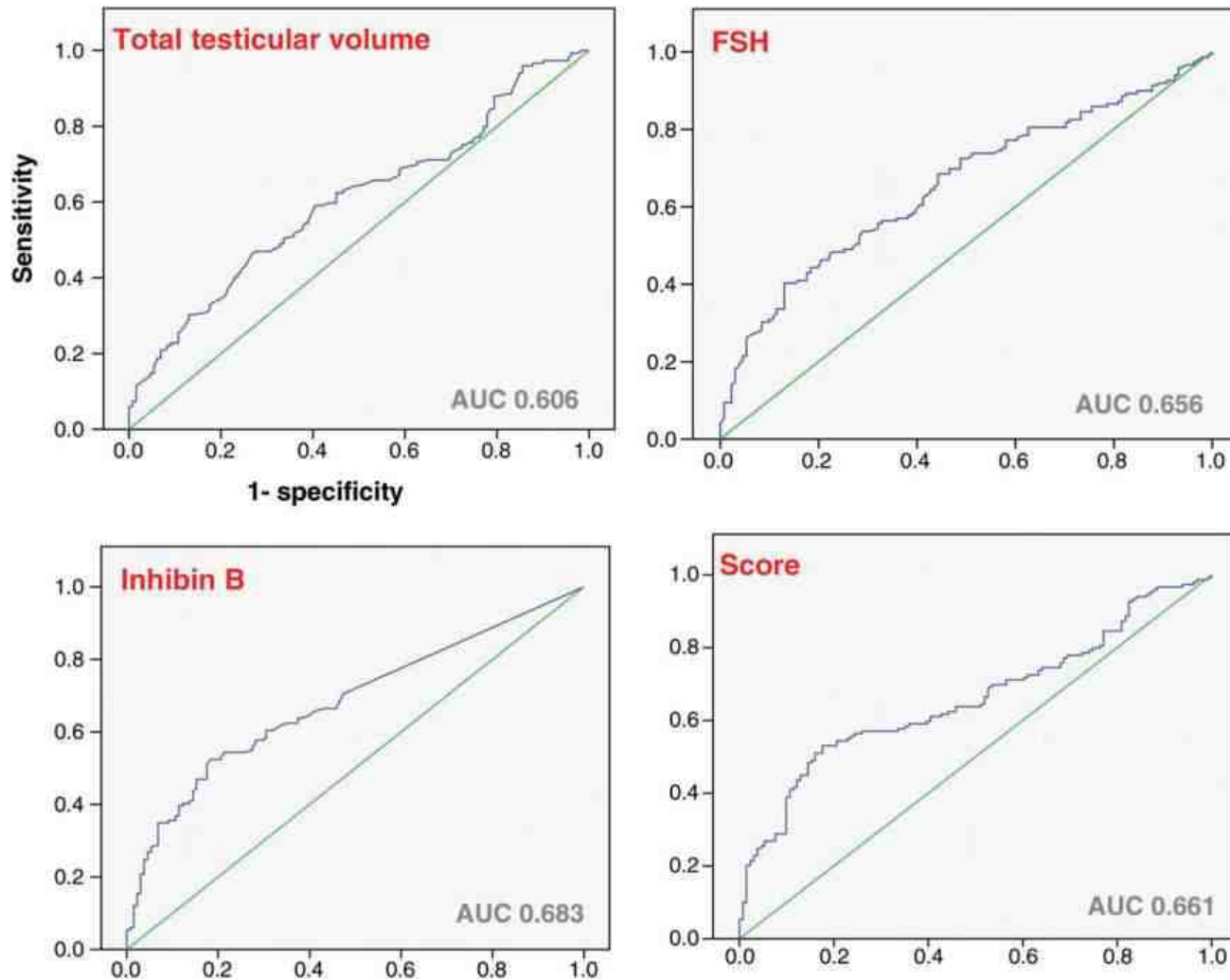
- ❑ Questionnaire of 530 children born after ICSI with testicular cells and 194 with epididymal sperm. Compared to 2516 with ejaculated cells
- ❑ Overall neonatal health in terms of birth parameters, major abnormalities and chromosomal aberrations was 'reassuring'

Predicting successful outcomes particularly for NOA (aside KS and Y deletions)

What screens?

- ❑ Traditionally techniques have limited usefulness (e.g. volume, FSH, inhibin B).
- ❑ Possibility of placing them together n= 280. Positive likelihood ratio~3.

ROC curves for TTV, FSH, inhibin B and our score as a guide to the presence of spermatozoa in 280 men with NOA.



Boitrelle F et al. Hum. Reprod. 2011;26:3215-3221

However...

- Still limited use but may help provide realistic chances for patients

Human Reproduction, Vol.26, No.12 pp. 3213–3214, 2011

Advanced Access publication on September 18, 2011 doi:10.1093/humrep/der316

human
reproduction

EDITORIAL COMMENTARY *Andrology*

How to predict fatherhood for men with non-obstructive azoospermia opting for TESE–ICSI?

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Potential new markers

- One example would be seminal plasma proteins

Research

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This paper is available on line at <http://www.mcponline.org>

Verification of Male Infertility Biomarkers in Seminal Plasma by Multiplex Selected Reaction Monitoring Assay*

Andrei P. Drabovich†, Keith Jarvi§¶, and Eleftherios P. Diamandis†§**††§§

Seminal plasma is a promising biological fluid to use for noninvasive clinical diagnostics of male reproductive system disorders. To verify a list of prospective male

essential diagnosis of azoospermia. *Molecular & Cellular Proteomics* 10: 10.1074/mcp.M110.004127, 1–13, 2011.

What types of germ cells from the testis can we inject? Any change?

Stages of mature spermatozoa?

Elongated spermatids?

Round spermatids?

How do we judge success?

Round Spermatis

ASRM (2008) summary

(ASRM Practice Committee Fertil Steril 90, Supplement 3 S199-200)

1. Generally unsuccessful – clinically used in rare circumstances
2. Experimental technique
3. Identification an issue
4. Oocyte activation
5. Genetic abnormalities 1 specific paper of concern
6. Poor embryonic development

Yana -Round spermatids may work.....

Table 1. Clinical outcome of ROSI partners of 76 women in this table had round spermatids as their most advanced spermatogenic cells

Assisted fertilization	Cryopreservation of embryos	Total no. of women	Total no. of treatment cycles	Percentage of oocytes fertilized (fert/total) (%)	Percentage Oocytes cleaved (cleaved/total) (%)	Total no. of transferred embryos	No. and percentage of recipients (%)				Total no. of live offspring
							Total transferred cycles	Pregnancy*	Miscarriage	Live offspring delivered	
ROSI (present study)	No	58	162	55.6 (330/594)	44.9 (267/594)	152	121	20 (16.5)	13 (65.0)	7 (5.8) ^{§§}	9
	Yes	28 [†]	42	76.4 (107/140)	71.4 (100/140)	56	42	10 (23.8)	5 (50.0)	5 (11.9) [¶]	5
TESE and ICSI	No	260	540	44.9 (833/1855)	43.5 (807/1,855)	562	468	134 (28.6)	27 (20.1)	107 (22.9) [§]	121
	Yes	140	295	68.2 (825/1210)	65.3 (790/1,210)	377	290	174 (60.0)	14 (8.0)	160 (55.2) ^{§§, ¶¶}	176

Partners of 76 women in this table had round spermatids as their most advanced spermatogenic cells. To compare the efficiencies of ROSI and ICSI, part of the data collected in our clinic after TESE and ICSI are included in this table. § vs. §§ and ¶ vs. ¶¶, $P < 0.05$; Fisher's protected least significant difference test.

*No and percentage of recipients with positive germinal sacs detected by ultrasound scanning between 4 and 5 wk of gestation.

[†]Ten of these 28 patients failed to get pregnancy previously when they received nonfrozen ROSI embryos.



Fourteen babies born after round spermatid injection into human oocytes

Atsushi Tanaka[§], Motori Nagayoshi[¶], Youichi Takemoto[¶], Izumi Tanaka[¶], Hiroshi Kusunoki[¶], Seiji Watanabe[¶], Keiji Kuroda[¶], Satoru Takeda[¶], Masahiko Ito[¶], and Ryuzo Yanagimachi^{§, ¶¶}

[§]Joint Member Obstetrics and Gynecology Clinic, Institute for ART, Tokyo 807-8075, Japan; [¶]Natural Diversity Science, Graduate School of Agriculture, Kinki University, Kobe 651-8501, Japan; ^{¶¶}Department of Anatomical Science, Mansai University Graduate School of Medicine, Hirumaki 330-8042, Japan; ^{¶¶}Department of Obstetrics and Gynecology, Juntendo University School of Medicine, Tokyo 113-8513, Japan; ^{¶¶}Department of Obstetrics, Osaka University School of Medicine, Yamoto-cho 4-1-176, Japan; and ^{¶¶}Department of Anatomy, Physiology, and Biochemistry, University of Texas Medical School, Houston, TX 77030

Contributed by Ryuzo Yanagimachi, September 25, 2015 (sent for review July 28, 2015); received by J. Michael Bedford and Martin M. Matzuk

Summary

Essentials and discussion points

- Physical examination and history
- **High Quality Semen Analyses**
- **If abnormal spermatogenesis : potential screens.**
 - **Genetics e.g. Y deletions**
 - **Routine ? FSH**
 - **Possible new seminal plasma**
 - **? Spermatids**

Repeated attempts can be successful.

Table II. Consecutive sperm retrieval in patients with nonobstructive azoospermia

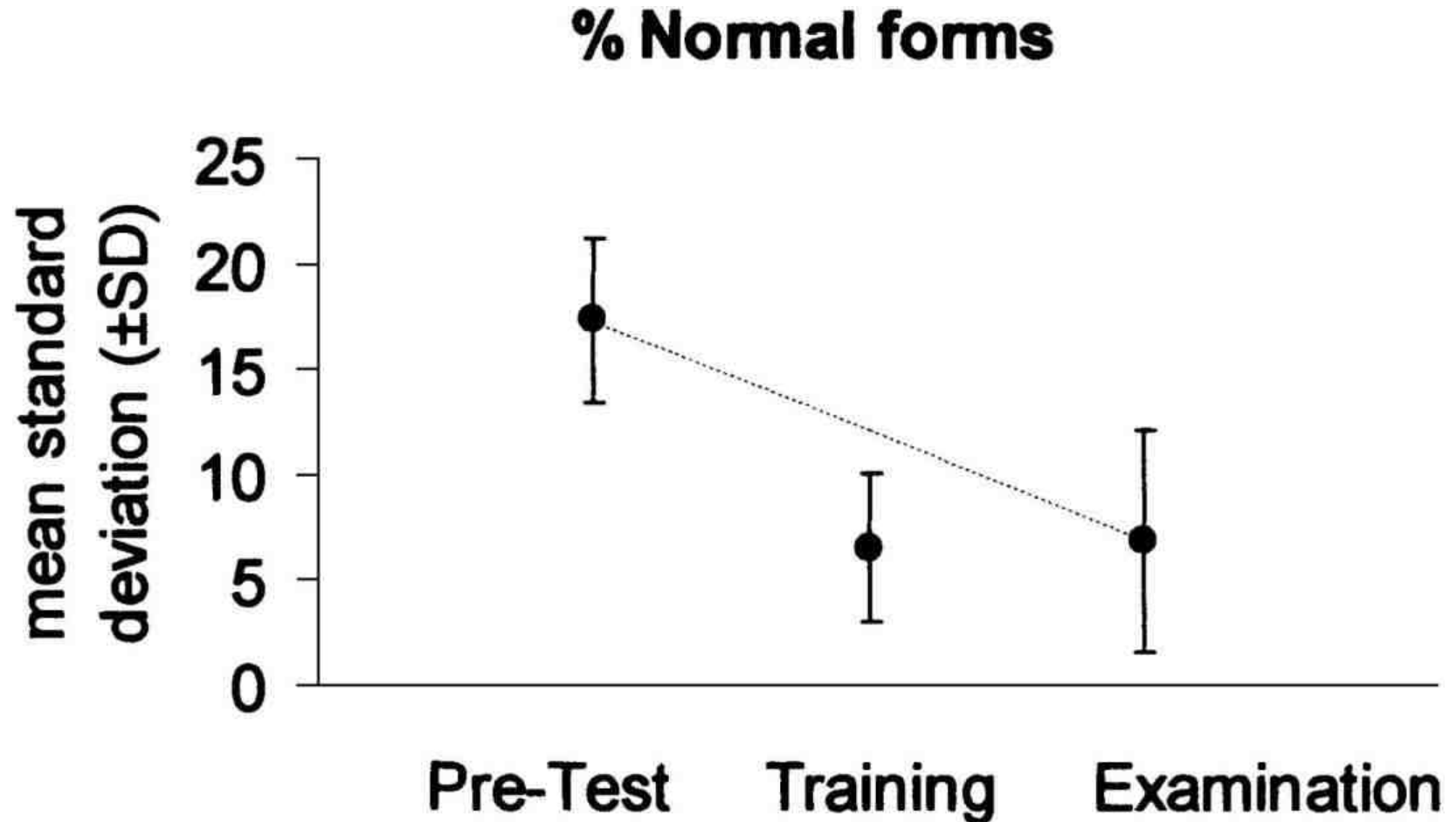
Rank	<i>n</i>	Successful sperm recovery (<i>n</i>)	Successful sperm recovery (%)
Total	784	384	49
1	628	261	41.6
2	103	77	74.7
3	34	28	82.3
4	11	11	100
5	6	5	83.3
6	2	2	100

From Vernaev et al., 2006
Hum Reprod 21, 1551-1554

Also issue of how to train and improved compliance

High quality consistent training does help

Decrease in course participants' variability, as revealed by average SD of results from assessments of proportion of morphologically normal sperm in test samples, from pre-test (n = 16), through training (n = 15) to examination (n = 15).



Björndahl L et al. Hum. Reprod. 2002;17:1299-1305

High quality consistent training does help

TABLE 1

Descriptive statistics of 290 participants' morphology scores during 18 pretraining and posttraining sessions.

	Normal cells recorded during pretraining (%)		Normal cells recorded during posttraining (%)	
	Slide 1	Slide 2	Slide 3	Slide 4
Reference slide value	15	5	14	9
Lowest value	0.0	0.0	0.0	-8.0
Highest value	98.0	75.0	60.0	33.0
Mean	29.7	12.6	13.2	4.4
95% confidence interval	26.6-33.2	10.5-14.7	12-14	3.4-5.4
Median	17.0	6.5	13.0	2.0
SD	26.6	16.2	8.0	5.6
Percent difference from mean	104.0 ± 189.1 ^{a,b}	13.0 ± 15.8 ^{c,d}	13.0 ± 8.1	4.2 ± 5.7

^a Compared with slide 3, $P = .0001$, Mann-Whitney test for independent samples.

^b Compared with slide 4, $P = .001$, Mann-Whitney test for independent samples.

^c Compared with slide 3, $P = \text{not significant}$, Mann-Whitney test for independent samples.

^d Compared with slide 4, $P = \text{not significant}$, Mann-Whitney test for independent samples.

Franken. Semenology training. *Fertil Steril* 2010.

Generally NOA and OA

- ❑ Cochrane meta analysis suggests no hard evidence to suggest one technique over another (van Peperstraten et al., (2008) Cochrane Database Syst Rev (16)CD002808)

- ❑ Men with spermatogenesis e.g. vasectomy reversal – possible from epididymis – motile sperm have similar success rates to testicular cells (Nicopoullos *et al.*, 2004 Fertil Steril 82, 691-701) and can be cryopreserved.

- ❑ NOA : TESE but maybe via microsurgery.
 - Recovery rates are very variable: 40-50% (Tournaye 2012 Asian J Andrology 14, 103-108)
 - Meta analysis: Deruyver Y, Vanderschueren D, Van der Aa F. Andrology. 2014 Jan;2(1):20-4)

EAU Guidelines

(Jungwirth et al., 2012 Eur Urol.
62:324-32)

Diagnosis	Unselected 12,945	Azoospermic 1446
Idiopathic	30.3 %	13.3%
KS (XXY)	2.6%	13.7%
Y deletions	0.3%	1.6%

Significant component unknown.