# ARTICLE IN PRESS

YGYNO-974269; No. of pages: 7; 4C:

Gynecologic Oncology xxx (2011) xxx-xxx



Contents lists available at SciVerse ScienceDirect

# **Gynecologic Oncology**

journal homepage: www.elsevier.com/locate/ygyno



#### Review

# The association between endometriosis and ovarian cancer: A review of histological, genetic and molecular alterations

Peter Svenssen Munksgaard, Jan Blaakaer\*

Department of Gynecology and Obstetrics, Aarhus University Hospital, Skejby, Denmark

#### ARTICLE INFO

Article history: Received 22 July 2011 Accepted 1 October 2011 Available online xxxx

Keywords: Endometriosis Ovarian cancer ARID1A Genetics

#### ABSTRACT

*Objective.* This article represents a review of histologic and genetic findings in endometriosis and describes the mechanisms whereby genetic and non-genetic factors potentially contribute to the neoplastic progression of endometriosis.

*Methods*. Literature review of the English language literature based on searching in the MEDLINE (PubMed) database and additional collection of reports by systematically reviewing all references from retrieved papers.

Results. Atypical endometriosis seems to represent a transition from benign endometriosis to carcinoma. Endometriosis is characterized by genetic instability: like neoplasms endometriosis seems to be monoclonal in origin, several studies have documented loss of heterozygosity (LOH) in endometriosis, data suggest that mutation of the tumor suppressor gene PTEN play a part in the malignant transformation of endometriosis, some studies have revealed TP53 mutations in endometriotic lesions, and mutation of ARID1A seems to be an important early event in the malignant transformation of endometriosis to endometrioid and clear cell carcinomas. Heme and iron induced oxidative stress, inflammation, and hyperestrogenism are possible links between endometriosis and cancer.

Conclusions. The histological and genetic alterations in endometriosis seem to explain why endometriosis can be a precursor of some ovarian cancers, especially clear cell and endometrioid carcinomas. However, the exact molecular mechanisms that may lead to this malignant transformation of endometriosis are not completely understood. More and larger studies are needed to clarify how exactly endometriotic tissue undergoes malignant transformation.

© 2011 Published by Elsevier Inc.

#### Contents

## Introduction

Endometriosis is an estrogen-dependent, chronic gynecological disorder and is classically defined as the presence of endometrial glands and stroma outside the uterine cavity and musculature [1,2]. The precise etiology of endometriosis is not known but one widely

0090-8258/\$ – see front matter © 2011 Published by Elsevier Inc. doi:10.1016/j.ygyno.2011.10.001

Please cite this article as: Munksgaard PS, Blaakaer J, The association between endometriosis and ovarian cancer: A review of histological, genetic and molecular alterations, Gynecol Oncol (2011), doi:10.1016/j.ygyno.2011.10.001

<sup>\*</sup> Corresponding author at: Department of Gynecology and Obstetrics, Aarhus University Hospital, Skejby, DK 8200 Aarhus N, Denmark. Fax: +45 8949 6024.

E-mail address: jab@dadlnet.dk (J. Blaakaer).

accepted mechanism for the development of endometriotic lesions is the adhesion and growth of endometrial fragments deposited into the peritoneal cavity via retrograde menstruation [3-5]. Although endometriosis is considered to be a benign condition, it does share common characteristics with malignant cells. Endometriosis, like cancer, can be both locally and distantly metastatic and it can attach to other tissues, invade, and damage them [6]. In 1925, Sampson [7] was the first to document the case of suspected malignant transformation in endometriosis. Since then, several studies have focused on the relationship between endometriosis and gynecological cancer, especially ovarian cancer. Data from large cohort and case-control studies [8–15] indicate that endometriosis patients have an increased risk of ovarian cancer. Further, endometriosis-associated ovarian cancers are predominantly clear cell and endometrioid histologic subtypes. That is, studies [8,12,15-29] have documented that endometriosis is associated with an approximately 3-fold statistically significant risk of endometrioid and clear cell ovarian cancer. In addition, endometriosis-associated ovarian cancer seems to be a distinct clinical entity; patients are younger, diagnosed in earlier stages, have lower grade lesions and a better survival [13,19,28-30].

In a recent paper we reviewed the epidemiological data concerning endometriosis and ovarian cancer [31]. The aim of this article is to represent a review of histologic and genetic findings in endometriosis and to describe the mechanisms whereby genetic and nongenetic factors potentially contribute to the neoplastic progression of endometriosis.

#### Methods

This article reviews the English language literature for studies on the histologic, genetic and molecular mechanisms that could potentially explain the increased risk of ovarian cancer among endometriosis patients. We searched in the MEDLINE (PubMed) database for a 20-year period (January 1990–June 2011), combining the keywords "endometriosis" with "neoplasms" or "ovarian neoplasms". Additional reports were collected by systematically reviewing all references from retrieved papers.

## Results

Atypical endometriosis: a precursor of malignancy

Similarly to the eutopic endometrium studies have documented the presence of hyperplasia and cytological atypia in endometriosis. Cytologic atypia denotes the presence of cytologic atypia within the lining of endometriotic cysts, whereas hyperplasia refers to the same spectrum of hyperplasia (simple or complex, with or without cytologic atypia) encountered in the endometrium. These two conditions are often referred to collectively as atypical endometriosis, but they should be separately recognized as their clinical significance likely differs [32]. Hence, the term "atypical" has been used with different meanings among pathologists. In general, several studies have suggested that atypical endometriosis may represent a transition from benign endometriosis to carcinoma [21,33–36].

LaGrenada and Silverberg [35] were the first to publish a case series of five ovarian tumors (three clear-cell and two endometrioid carcinomas) associated with atypical endometriosis. In four of these tumors a direct continuous transition was seen from clearly benign epithelium through atypical endometriosis to carcinoma. A case report by Moll et al. [36] demonstrated a chronological association; a woman who on a biopsy specimen first showed atypia within ovarian endometriosis, and then 3 years later had a clear cell carcinoma arising in the same ovary. In a review of 339 patients with cystic ovarian endometriosis Prefumo et al. [34] found that atypia was significantly more present in patients with endometriosis-associated ovarian cancer (endometrioid adenocarcinoma) compared to patients with

endometriosis alone (100% of 14 cases compared to 2% of 325 controls, P<0.00001). Further, the authors documented that complex hyperplasia, but not simple hyperplasia, was significantly more common in cases with associated ovarian carcinoma than in those without malignancy (50% of 14 compared to 1% of 325, P<0.00001). This is supported by Fukunaga et al. [33], who reported that atypical endometriotic foci were observed in 61% of 54 endometriosis-associated ovarian cancers, while such foci were seen in only 1.7% of 255 endometriosis cases without cancers. Finally, Ogawa et al. [21] observed atypical endometriosis in 29 out of 37 (78%) cases with endometriosis-associated ovarian cancer. Interestingly, the transition from typical endometriosis to atypical endometriosis was observed in 22 cases, and the transition from atypical endometriosis to carcinoma, in 23 cases.

Given this evidence atypical endometriosis (both cytological atypia and hyperplasia) is likely a lesion that represents a transition from benign endometriosis to carcinoma.

Genetic alterations in endometriotic tissue

Genomic instability is a known characteristic of cancer cells [37]. In addition, it is well known that most neoplasms are monoclonal in origin, and clonal outgrowth is thought to be a fundamental feature of human neoplasms [38,39]. Several studies have demonstrated evidence for monoclonality of endometriosis. In seven studies [40–46] identified from 1995 to 2003 endometriotic tissue was monoclonal in 60–100% of the samples investigated (100% in five of the studies). However, these findings have been challenged by Mayr et al. [47], who only found monoclonality in 2 of 32 (6%) samples of endometriotic lesions.

Other studies have documented loss of heterozygosity (LOH) in endometriosis [41,48-52]. LOH commonly indicates regions of tumor suppressor gene inactivation, which is central to the development of malignant tumors as well as their precursors. Jiang et al. [41] revealed LOH at candidate ovarian tumor suppressor gene loci in 11 of 40 (27.5%) endometriosis samples. In another study [48], the same authors detected common LOH events in 9 of 11 cases of ovarian endometriosis adjacent or contiguous to ovarian carcinoma. Based on statistical analysis, these common genetic alterations were very unlikely to have occurred as independent events, indicating a possible malignant genetic transition spectrum between endometriosis and cancer. This has been further supported by Prowse et al. [49], who analyzed four endometrioid and six clear cell carcinomas with coexisting endometriosis and found 22 common LOH events. Again, statistical analyses suggested that it was very unlikely that these common genetic alterations had occurred independently. In addition, Obata et al. [50] observed frequent LOH in ovarian atypical endometriosis, and Goumenou et al. [51] found LOH at loci harboring candidate genes implicated in limiting malignant transformation in 8 of 22 (36.4%) endometriosis samples. Interestingly, occurrence of these genetic alterations increased 3-fold from stage II ovarian endometriosis to more severe stages of the disease, suggesting that LOH events accumulate as the disease progresses.

LOH at locus 10q23.3 and mutation of the tumor suppressor gene PTEN (located on 10q23.3) occur frequently in both endometrial and ovarian carcinoma. Sato et al. [52] identified LOH at 10q23.3 in 13 of 23 of endometrial cysts of the ovary (56.5%), 8 of 19 ovarian endometrioid carcinomas (42.1%), and 6 of 22 clear cell carcinomas (27.3%). Further, somatic mutations in the PTEN gene were identified in 20.6% of endometrial cysts of the ovary, 20% of endometrioid carcinomas and 8.3% of clear cell carcinomas. This suggests that inactivation of the PTEN is an early event in the malignant transformation of endometriosis. These findings are supported by Martini et al. [53], who detected reduced PTEN protein expression in 7 of 46 (15%) cases of endometriosis. In addition, a mouse model by Dinulescu et al. [54] provided a support for a role of PTEN in the malignant transformation

P.S. Munksgaard, J. Blaakaer / Gynecologic Oncology xxx (2011) xxx-xxx

of endometriosis; forced expression of oncogenic K-ras or conditional PTEN deletion in ovarian surface epithelium resulted in preneoplastic ovarian lesions with an endometrial glandular morphology, while the combination of both gave rise to invasive and widely metastatic endometrioid ovarian adenocarcinomas. In conclusion, the described LOH events represent a convincing piece of evidence that there might be a continuum between endometriosis and ovarian cancer, the last-mentioned events being the most important. The results from the LOH studies are summarized in Table 1.

Mutation or loss of function of the tumor suppressor gene TP53 (on chromosome 17p13.1), which encodes for the nuclear protein p53, is a frequent and important genetic event in the development of ovarian cancer. Mutations of the TP53 gene are related to overexpression of non-functional p53 protein, which accumulates within the nuclei [55,56]. Therefore, several studies have investigated whether endometriosis harbor mutations in this gene [41,57-62]. Bischoff et al. [57] reported that chromosomal loss of TP53 frequently occurred in severe/late stage endometriosis. Nezhat et al. [58] detected p53 accumulation in 9% and 25% of benign endometriotic lesions next to clear cell carcinoma and endometrioid carcinoma, respectively, while p53 staining was negative in endometriosis without carcinoma, Further, Sainz de la Cuesta et al. [59] found a statistically significant prevalence of p53 overexpression in the transition from typical to atypical endometriosis and to ovarian cancer associated with endometriosis. Finally, Akahane et al. [60] observed TP53 mutation in 4 of 13 (30.8%) specimens of endometriosis coexisting with ovarian clear cell carcinoma, whereas this study did not detect mutations in seven cases of solitary ovarian endometriosis and nine cases of endometriosis coexisting with ovarian endometrioid carcinoma. Moreover, other studies [41,61,62] have also failed to detect mutations in the TP53 gene and/or p53 overexpression in solitary endometriotic lesions not associated with carcinomas. In addition, several published studies show no or very low percentage of TP53 mutation in clear cell ovarian cancer [63,64].

Most recently, a very interesting study by Wiegand et al. [65] has identified ARID1A mutations in endometriosis-associated ovarian carcinomas (clear-cell and endometrioid carcinomas). ARID1A is thought to be a tumor-suppressor gene, and the protein encoded by the ARID1A, BAF250a, is a key component of the multi-protein SWI/ SNF chromatin remodeling complex present in all eukaryotes. This complex is involved in the regulation of many cellular processes, including development, differentiation, proliferation, DNA-repair, and tumor suppression. Based on genome-wide mutational analysis, ARID1A mutations were seen in 55 of 119 ovarian clear-cell carcinomas (46%), 10 of 33 endometrioid carcinomas (30%), and none of the 76 high-grade serous ovarian carcinomas. Immunohistochemical analyses confirmed the loss of expression of the BAF250a protein and its correlation with AIRD1A mutation status. More important, in two patients, AIRD1A mutations and loss of BAF250a expression were evident in the tumor and contiguous atypical endometriosis but not in distant endometriotic lesions. These results provide convincing evidence for the inactivation of ARID1A in the development of clear-cell and endometrioid ovarian cancers. Further, the data suggest that ARID1A inactivation occurs early in the development of these tumors, during transformation of endometriosis. In addition, the presence of ARID1A mutations in ovarian clear cell and uterine endometrioid carcinomas have also been documented by Jones et al. [66] and Guan et al. [67], respectively.

Heme and free iron induced oxidative stress

Heme and free iron are pro-oxidant and can induce oxidative stress and DNA damage, possibly increasing the risk of some cancer [68,69]. Retrograde menstruation, a widely accepted etiology of endometriosis, and the repetition of hemorrhage in the endometriotic (chocolate) cysts during the reproductive period result in excess accumulation of heme and free iron in endometriotic lesions, which may play a role in carcinogenesis [69,70]. Yamaguchi et al. [70] found that the free iron concentration in endometriotic cysts was significantly higher than that in nonendometriotic cysts (P<0.01). In addition, the concentrations of stress-related factors such as lactose dehydrogenase (a marker of tissue damage), lipid peroxidase (a marker of oxidative stress), and 8-hydroxy-2-deoxyguanosine (a marker of DNA damage) were also significantly higher in endometriotic cysts. Moreover, in vitro analyses showed that the contents of endometriotic cysts could produce more reactive oxygen species and induce gene mutations more frequently than the contents in the other cysts.

#### Inflammation

As noted for centuries, inflammation may be central to carcinogenesis. As described by Balkwill and Mantovani [71] "If genetic damage is 'the match that lights the fire' of cancer, some types of inflammation may provide the 'fuel that feeds the flames'". Inflammatory cells and cytokines may promote angiogenesis, cell proliferation, inhibition of apoptosis, invasion, metastasis, and production of reactive oxygen species that may in turn induce DNA damage and mutations. Thus, inflammation may contribute to tumor growth and progression [71,72].

Endometriosis is associated with a local inflammatory reaction; inflammatory cells surround sites of endometriosis, and increased levels of cytokines and growth factors have been observed in the peritoneal fluid of women with endometriosis [73,74]. It has been proposed that this inflammation may serve to promote the growth and invasion of ectopic endometrium and constitute a link between endometriosis and ovarian cancer [75]. As an example, a study by Daraï et al. [76] reported that endometriosis was associated with serum tumor necrosis factor-alpha levels similar to those found in women with ovarian cancer, while serum interleukin-6 levels and cyst fluid interleukin-8 levels were intermediate between those observed in benign and malignant ovarian tumors. Slater et al. [77] observed that levels of growth hormone and interleukin-6 were increased in endometriosis and endometrioid adenocarcinomas, compared to normal uterine epithelial cells, suggesting that these proteins appear to be involved in the progression of both conditions. High concentrations of interleukin-1 have been detected in the peritoneal fluid of women with endometriosis [74]. In addition, Wu et al. [78] found that ectopic endometrium was at least 100 times more sensitive to interleukin-1beta (IL-1beta), compared with eutopic endometrium. IL-1beta can upregulate cyclooxygenase-2 (COX-2) expression, leading to subsequent increased synthesis of prostaglandin E2 (PGE2). Interestingly, PGE2 regulates key-processes, which are characteristic for

 Table 1

 LOH events in endometriosis, ovarian clear cell carcinomas (CCC), ovarian endometrioid carcinomas (EC) and common events in coexisting endometriosis and ovarian cancer.

Author	Endometriosis (%)	CCC (%)	EC (%)	Common LOH events in coexisting endometriosis and ovarian cancer (%)
Jiang et al. 1996 [41] Jiang et al. 1998 [48] Prowse et al. 2006 [49] Goumenou et al. 2001 [51]	27.5			81,8 22 common LOH events in 4 EC and 6 CCC with coexisting endometriosis
Sato et al. 2000 [52]	56.5	27.3	42.1	

Please cite this article as: Munksgaard PS, Blaakaer J, The association between endometriosis and ovarian cancer: A review of histological, genetic and molecular alterations, Gynecol Oncol (2011), doi:10.1016/j.ygyno.2011.10.001

tumor growth, e.g. angiogenesis, proliferation, and inhibition of apoptosis [79,80]. Moreover, Rask et al. [81] have demonstrated an increase of COX-2 in epithelial ovarian cancer, correlating to stage with significant elevations in stages II and III.

#### Steroid hormones

Estrogens have been linked to the pathogenesis of the gynecological cancers; breast-, endometrium-, and ovary-cancer [82–84]. Moreover, hyperestrogenism stimulates the growth of endometriosis and seems to be associated with the malignant transformation of endometriotic cysts [85,86]. Some studies [87,88] have demonstrated a local increase in estradiol concentration in endometriotic lesions. The enzyme aromatase catalyzes the conversion of androstenedione and testosterone, derived from ovarian and adrenal sources, to estrone and estradiol, respectively. This enzyme is normally absent in eutopic endometrium, but studies by Zeitoun and Bulun [87] have shown high levels of aromatase activity in endometriotic lesions, resulting in increased conversion of androgens to estrogens. Further, another study by Zeitoun et al. [88] found that the enzyme (17betahydroxysteroid dehydrogenase type 2), which catalyzes the inactivation of estradiol to the less potent estrone, was absent in endometriotic lesions but was normally expressed in eutopic endometrium. In addition, the enzyme (17beta-hydroxysteroid dehydrogenase type 1) converting estrone to estradiol was present in endometriosis. In sum, synthesis of the potent estradiol is increased while inactivation is decreased resulting in higher local concentrations of this hormone. Estradiol stimulates cyclooxygenase-2 (COX-2), giving rise to increased PGE2 formation. PGE2 is as mentioned above implicated in tumor progression but also stimulates further aromatase activity creating a positive feedback loop in favor of continuous estrogen formation in endometriosis [87]. A study by O'Donnell et al. [89] has shown that the potential carcinogenic action of estrogens in ovarian cancer cells was mediated through estrogen receptor alpha (ER-alpha), whereas estrogen receptor beta (ER-beta) did not seem to be involved. Interestingly, Matsuzagi et al. [90] have demonstrated higher ER-alpha than ER-beta levels in active endometriosis compared to inactive endometriosis, suggesting that ER-alpha may be essential for the pathogenesis of endometriosis.

Progesterones generally repress estrogen receptors, inhibiting the effects of estrogens at the cellular level. However, there is evidence of a relative progesterone resistance within endometriotic lesions; the presence of the inhibitory progesterone receptor isoform A, the absence of the stimulatory progesterone receptor isoform B, and progesterone receptor gene polymorphism have been observed in endometriosis [2,91,92]. This could lead to further escalation of estrogen actions on endometriotic lesions.

### Discussion

Generally speaking, support to the idea of endometriosis as a precursor of some ovarian cancers comes from the cases of atypical endometriosis with evidence of a histologically-proven transition from benign disease to malignancy. However, conclusive evidence is lacking on this subject, and it cannot be ruled out that the occurrence of atypical endometriosis in ovarian carcinomas is purely coincidental. Further, if ovarian carcinomas do arise from endometriosis, one would expect to find genetic alterations common to both diseases. Conversely, if endometriosis represents a distinct entity then one would expect to find alterations unique to this disease. The genetic studies tend to suggest that endometriotic lesions may harbor mutations in genes critical to cancer development. Moreover, the frequency of genetic aberrations seems to increase for cases of endometriosis adjacent or contiguous to ovarian cancer, and these cases tend to share the same genetic mutations, which supports the model of endometriosis as a malignant precursor. However, it is not clear whether

these genetic alterations support causality as poor consistency exists among the DNA analyses of endometriosis, and the real frequency of these genetic alterations is presently unknown [93]. The recent study by Wiegand et al. [65] may have found a definitive link suggesting that clear cell and endometrioid carcinomas arise from endometriosis and that mutation of ARID1A is important in this process. However, this conclusion is based on only two observations, necessitating extensive validation involving endometriotic lesions from patients with ovarian cancer and patients without cancer. As mentioned above ARID1A mutations in uterine carcinomas have also been documented [67,94]. This is not surprising and further supports the hypothesis of endometriosis as a malignant precursor, since clear cell and endometrioid carcinomas of the ovary seem to be derived from ectopic uterine epithelium, that is, endometriosis. It also raises the possibility that treatments that are effective for uterine clear cell and endometrioid carcinomas could be effective for their ovarian counterparts. Furthermore, if the loss of ARID1A is an early event in the malignant transformation of endometriosis, then perhaps this intervention could serve as a prevention strategy. Should endometriotic lesions be analyzed for expression of BAF250a, the protein encoded by ARID1A? Should patients with endometriotic lesions that show loss of expression be viewed as being at high risk for the development of clear-cell or endometrioid ovarian cancers? Much work remains to be done to define the role of ARID1A in the origin of ovarian cancer. Nonetheless, the discovery by Wiegand et al. has provided an important step forward and the origin of ovarian cancer is becoming clearer.

Recently, our knowledge of epithelial ovarian cancer has evolved decisively. Molecular genetic studies [63,64,95] have led to the development of a new paradigm for the pathogenesis and origin of epithelial ovarian cancer based on a dualistic model of carcinogenesis that divides epithelial ovarian cancer into two broad categories designated types 1 and 2. Type 1 tumors are composed of low-grade serous, low grade endometrioid, clear cell, mucinous, and transitional (Brenner) carcinomas. They are genetically indolent, present in stage 1 and the majority is characterized by specific mutations, including PTEN and ARID1A. Though belonging to type 1 low grade tumor clear cell and mucinous tumors are generally chemoresistant and they are quite aggressive particularly at late stages. Type 2 tumors comprise high grade serous, high-grade endometrioid, malignant mixed mesodermal tumors, and undifferentiated carcinomas. They are aggressive, present in advanced stage, and have a very high frequency of TP53 mutations but rarely harbor the mutations detected in type 1 tumors. In addition, it has been proposed that serous tumors arise from the implantation of epithelium (benign or malignant) from the fallopian tube. Similarly, it seems that endometriosis is the precursor of endometrioid and clear cell carcinomas. As endometriosis is thought to develop from endometrial tissue by retrograde menstruation, these tumors can also be regarded as involving the ovary secondarily. Thus, it appears that both type 1 and 2 ovarian tumors develop independently along different molecular pathways and that both types develop outside the ovary and involve it secondarily. This new dualistic model can explain why some studies [41,60–62] have failed to detect mutations in the TP53 gene and/or p53 overexpression in endometriotic lesions because this mutation is only associated with high grade endometrioid carcinomas and not the other endometriosis-associated ovarian cancers, low-grade endometrioid and clear cell carcinomas. Overall, these findings represent a major shift in our understanding of the origins of ovarian carcinomas and will allow for a more rational approach to screening, treatment, and prevention that potentially can have a significant impact on reducing the mortality of this devastating disease. For example, the heterogeneity of ovarian carcinoma clearly indicates that one screening test will not be effective in detecting all the different types of ovarian carcinomas. In addition, the genes in types 1 and 2 could provide potential targets for therapeutic intervention and moreover, future analysis of overexpressed genes in ovarian cancer should take into account the histologic type of the tumors being studied and the data should be compared with appropriate normal tissue.

In conclusion, the exact molecular mechanisms that may lead to malignant transformation of endometriosis are not completely understood. As regards heme and iron induced oxidative stress, inflammation, and hyperestrogenism, these theories are possible links between endometriosis and cancer, but definitive data are lacking. More and larger studies are needed to address this problem. Yet, the studies imply interesting knowledge and hormonal modulation, antioxidant, and anti-inflammatory agents could provide future prevention and treatment of endometriosis-mediated lesions. Today, progestin therapy is a standard treatment for endometriosis but as mentioned above studies [2,91,92] have demonstrated a relative progesterone resistance within endometriotic lesions. This finding could explain why endometriosis in some patients fails to regress in response to treatment with progestins. The potential etiology regarding the association between endometriosis and ovarian cancer seems to be multifactorial. For example, clear cell and endometrioid ovarian carcinomas develop from endometriosis that frequently involves multiple sites in the pelvis, but still these neoplasms are almost always confined to the ovaries. It is likely that the propensity for growth in the ovary is multifactorial, but the precise reasons for this are unknown [63]. Given the high prevalence of endometriosis and the high mortality of ovarian cancer the potential association has important public health implications. Identification and characterization of somatic mutations and possible carcinogenic factors such as heme and iron induced oxidative stress, inflammation, and hyperestrogenism are not only fundamental in understanding the molecular pathogenesis of cancer but also can provide the rationale for the development of personalized diagnostic tests and therapy, and that is why it is essential to further investigate the histologic, genetic and molecular alterations in endometriotic tissue.

#### Conflict of interest statement

The authors declare that there are no conflicts of interest.

#### References

- [1] Olive DL, Schwartz LB. Endometriosis. N Engl J Med Jun 17 1993;328:1759-69.
- [2] Giudice LC, Kao LC. Endometriosis. Lancet Nov 13–19 2004;364:1789–99.
- [3] Gazvani R, Templeton A. New considerations for the pathogenesis of endometriosis. Int J Gynaecol Obstet Feb 2002;76:117–26.
- [4] Sampson J. Peritoneal endometriosis due to menstrual dissemination of endometrial tissue into the peritoneal cavity. Am J Obstet Gynecol 1927;14:442–69.
- [5] Halme J, Hammond MG, Hulka JF, Raj SG, Talbert LM. Retrograde menstruation in healthy women and in patients with endometriosis. Obstet Gynecol Aug 1984;64: 151–4.
- [6] Vlahos NF, Economopoulos KP, Fotiou S. Endometriosis, in vitro fertilisation and the risk of gynaecological malignancies, including ovarian and breast cancer. Best Pract Res Clin Obstet Gynaecol Feb 2010;24:39–50.
- [7] Sampson J. Endometrial carcinoma of the ovary, arising in endometrial tissue in that organ. Arch Surg 1925;10:1–72.
- [8] Brinton LA, Sakoda LC, Sherman ME, Frederiksen K, Kjaer SK, Graubard BI, et al. Relationship of benign gynecologic diseases to subsequent risk of ovarian and uterine tumors. Cancer Epidemiol Biomarkers Prev Dec 2005;14:2929–35.
- [9] Kobayashi H, Sumimoto K, Moniwa N, Imai M, Takakura K, Kuromaki T, et al. Risk of developing ovarian cancer among women with ovarian endometrioma: a cohort study in Shizuoka, Japan. Int J Gynecol Cancer Jan–Feb 2007;17:37–43.
- [10] Olson JE, Cerhan JR, Janney CA, Anderson KE, Vachon CM, Sellers TA. Postmenopausal cancer risk after self-reported endometriosis diagnosis in the Iowa Women's Health Study. Cancer Mar 1 2002;94:1612–8.
- [11] Ness RB, Grisso JA, Cottreau C, Klapper J, Vergona R, Wheeler JE, et al. Factors related to inflammation of the ovarian epithelium and risk of ovarian cancer. Epidemiology Mar 2000:11:111–7.
- [12] Ness RB, Cramer DW, Goodman MT, Kjaer SK, Mallin K, Mosgaard BJ, et al. Infertility, fertility drugs, and ovarian cancer: a pooled analysis of case–control studies. Am J Epidemiol Feb 1 2002;155:217–24.
- [13] Borgfeldt C, Andolf E. Cancer risk after hospital discharge diagnosis of benign ovarian cysts and endometriosis. Acta Obstet Gynecol Scand Apr 2004;83: 395–400.
- [14] Modugno F, Ness RB, Allen GO, Schildkraut JM, Davis FG, Goodman MT. Oral contraceptive use, reproductive history, and risk of epithelial ovarian cancer in women with and without endometriosis. Am J Obstet Gynecol Sep 2004;191: 733–40.

- [15] Rossing MA, Cushing-Haugen KL, Wicklund KG, Doherty JA, Weiss NS. Risk of epithelial ovarian cancer in relation to benign ovarian conditions and ovarian surgery. Cancer Causes Control Dec 2008:19:1357-64.
- [16] Kobayashi H, Sumimoto K, Kitanaka T, Yamada Y, Sado T, Sakata M, et al. Ovarian endometrioma—risks factors of ovarian cancer development. Eur J Obstet Gynecol Reprod Biol Jun 2008:138:187–93.
- [17] Vercellini P, Parazzini F, Bolis G, Carinelli S, Dindelli M, Vendola N, et al. Endometriosis and ovarian cancer. Am | Obstet Gynecol |ul 1993;169:181–2.
- [18] Sainz de la Cuesta R, Eichhorn JH, Rice LW, Fuller Jr AF, Nikrui N, Goff BA. Histologic transformation of benign endometriosis to early epithelial ovarian cancer. Gynecol Oncol Feb 1996:60:238–44
- [19] Toki T, Fujii S, Silverberg S. A clinicopathologic study on the association of endometriosis and carcinoma of the ovary using a scoring system. Int J Gynecol Cancer 1996:6:68–75.
- [20] Jimbo H, Yoshikawa H, Onda T, Yasugi T, Sakamoto A, Taketani Y. Prevalence of ovarian endometriosis in epithelial ovarian cancer. Int J Gynaecol Obstet Dec 1997:59:245–50.
- [21] Ogawa S, Kaku T, Amada S, Kobayashi H, Hirakawa T, Ariyoshi K, et al. Ovarian endometriosis associated with ovarian carcinoma: a clinicopathological and immunohistochemical study. Gynecol Oncol May 2000;77:298–304.
- [22] Vercellini P, Scarfone G, Bolis G, Štellato G, Carinelli S, Crosignani PG. Site of origin of epithelial ovarian cancer: the endometriosis connection. BJOG Sep 2000;107: 1155–7
- [23] Takahashi K, Kurioka H, Irikoma M, Ozaki T, Kanasaki H, Miyazaki K. Benign or malignant ovarian neoplasms and ovarian endometriomas. J Am Assoc Gynecol Laparosc May 2001;8:278–84.
- [24] Oral E, Ilvan S, Tustas E, Korbeyli B, Bese T, Demirkiran F, et al. Prevalence of endometriosis in malignant epithelial ovary tumours. Eur J Obstet Gynecol Reprod Biol Jul 1 2003;109:97–101.
- [25] Deligdisch L, Penault-Llorca F, Schlosshauer P, Altchek A, Peiretti M, Nezhat F. Stage I ovarian carcinoma: different clinical pathologic patterns. Fertil Steril Oct 2007;88: 906–10.
- [26] Stern RC, Dash R, Bentley RC, Snyder MJ, Haney AF, Robboy SJ. Malignancy in endometriosis: frequency and comparison of ovarian and extraovarian types. Int J Gynecol Pathol Apr 2001;20:133–9.
- [27] Kontoravdis A, Augoulea A, Lambrinoudaki I, Christodoulakos G, Tzortziotis D, Grammatikakis I, et al. Ovarian endometriosis associated with ovarian cancer and endometrial-endocervical polyps. J Obstet Gynaecol Res Jun 2007;33:294–8.
- [28] Erzen M, Rakar S, Klancnik B, Syrjanen K. Endometriosis-associated ovarian carcinoma (EAOC): an entity distinct from other ovarian carcinomas as suggested by a nested case–control study. Gynecol Oncol Oct 2001;83:100–8.
- [29] Modesitt SC, Tortolero-Luna G, Robinson JB, Gershenson DM, Wolf JK. Ovarian and extraovarian endometriosis-associated cancer. Obstet Gynecol Oct 2002;100: 788–95.
- [30] McMeekin DS, Burger RA, Manetta A, DiSaia P, Berman ML. Endometrioid adenocarcinoma of the ovary and its relationship to endometriosis. Gynecol Oncol Oct 1995;59:
- [31] Munksgaard PS, Blaakaer J. The association between endometriosis and gynecological cancers and breast cancer: a review of epidemiological data. Gynecol Oncol Oct 2011;123:157–63.
- [32] Clement PB. The pathology of endometriosis: a survey of the many faces of a common disease emphasizing diagnostic pitfalls and unusual and newly appreciated aspects. Adv Anat Pathol Jul 2007;14:241–60.
- [33] Fukunaga M, Nomura K, Ishikawa E, Ushigome S. Ovarian atypical endometriosis: its close association with malignant epithelial tumours. Histopathology Mar 1997;30:249–55.
- [34] Prefumo F, Todeschini F, Fulcheri E, Venturini PL. Epithelial abnormalities in cystic ovarian endometriosis. Gynecol Oncol Feb 2002;84:280–4.
- [35] LaGrenade A, Silverberg SG. Ovarian tumors associated with atypical endometriosis. Hum Pathol Sep 1988;19:1080–4.
- [36] Moll UM, Chumas JC, Chalas E, Mann WJ. Ovarian carcinoma arising in atypical endometriosis. Obstet Gynecol Mar 1990;75:537–9.
- [37] Hanahan D, Weinberg RA. The hallmarks of cancer. Cell Jan 7 2000;100:57–70.
- [38] Fialkow PJ. Clonal Origin of human tumors. Biochim Biophys Acta Oct 12 1976;458:283–321.
- [39] Nowell PC. The clonal evolution of tumor cell populations. Science Oct 1 1976;194: 23–8.
- [40] Nilbert M, Pejovic T, Mandahl N, Iosif S, Willen H, Mitelman F. Monoclonal origin of endometriotic cysts. Int J Gynecol Cancer Jan 1995;5:61–3.
- [41] Jiang X, Hitchcock A, Bryan EJ, Watson RH, Englefield P, Thomas EJ, et al. Microsatellite analysis of endometriosis reveals loss of heterozygosity at candidate ovarian tumor suppressor gene loci. Cancer Res Aug 1 1996;56:3534–9.
- [42] Jimbo H, Hitomi Y, Yoshikawa H, Yano T, Momoeda M, Sakamoto A, et al. Evidence for monoclonal expansion of epithelial cells in ovarian endometrial cysts. Am J Pathol Apr 1997;150:1173–8.
- [43] Tamura M, Fukaya T, Murakami T, Uehara S, Yajima A. Analysis of clonality in human endometriotic cysts based on evaluation of X chromosome inactivation in archival formalin-fixed, paraffin-embedded tissue. Lab Invest Feb 1998;78: 213–8.
- [44] Jimbo H, Hitomi Y, Yoshikawa H, Yano T, Momoeda M, Yasugi T, et al. Clonality analysis of bilateral ovarian endometrial cysts. Fertil Steril Dec 1999;72: 1142–3.
- [45] Wu Y, Basir Z, Kajdacsy-Balla A, Strawn E, Macias V, Montgomery K, et al. Resolution of clonal origins for endometriotic lesions using laser capture microdissection and the human androgen receptor (HUMARA) assay. Fertil Steril Mar 2003;79(Suppl. 1): 710-7.

- [46] Nabeshima H, Murakami T, Yoshinaga K, Sato K, Terada Y, Okamura K. Analysis of the clonality of ectopic glands in peritoneal endometriosis using laser microdissection. Fertil Steril Nov 2003;80:1144–50.
- [47] Mayr D, Amann G, Siefert C, Diebold J, Anderegg B. Does endometriosis really have premalignant potential? A clonal analysis of laser-microdissected tissue. FASEB J Apr 2003;17:693–5.
- [48] Jiang X, Morland SJ, Hitchcock A, Thomas EJ, Campbell IG. Allelotyping of endometriosis with adjacent ovarian carcinoma reveals evidence of a common lineage. Cancer Res Apr 15 1998:58:1707–12.
- [49] Prowse AH, Manek S, Varma R, Liu J, Godwin AK, Maher ER, et al. Molecular genetic evidence that endometriosis is a precursor of ovarian cancer. Int J Cancer Aug 1 2006;119:556–62
- [50] Obata K, Hoshiai H. Common genetic changes between endometriosis and ovarian cancer. Gynecol Obstet Invest 2000;50(Suppl. 1):39–43.
- [51] Goumenou AG, Arvanitis DA, Matalliotakis IM, Koumantakis EE, Spandidos DA. Microsatellite DNA assays reveal an allelic imbalance in p16(Ink4), GALT, p53, and APOA2 loci in patients with endometriosis. Fertil Steril Jan 2001;75:160–5.
- [52] Sato N, Tsunoda H, Nishida M, Morishita Y, Takimoto Y, Kubo T, et al. Loss of heterozygosity on 10q23.3 and mutation of the tumor suppressor gene PTEN in benign endometrial cyst of the ovary: possible sequence progression from benign endometrial cyst to endometrioid carcinoma and clear cell carcinoma of the ovary. Cancer Res Dec 15 2000;60:7052–6.
- [53] Martini M, Ciccarone M, Garganese G, Maggiore C, Evangelista A, Rahimi S, et al. Possible involvement of hMLH1, p16(INK4a) and PTEN in the malignant transformation of endometriosis. Int J Cancer Dec 1 2002;102:398–406.
- [54] Dinulescu DM, Ince TA, Quade BJ, Shafer SA, Crowley D, Jacks T. Role of K-ras and Pten in the development of mouse models of endometriosis and endometrioid ovarian cancer. Nat Med Jan 2005;11:63–70.
- 55] Schuijer M, Berns EM. TP53 and ovarian cancer. Hum Mutat Mar 2003;21:285–91.
- [56] Hogdall EV, Christensen L, Hogdall CK, Frederiksen K, Gayther S, Blaakaer J, et al. Distribution of p53 expression in tissue from 774 Danish ovarian tumour patients and its prognostic significance in ovarian carcinomas. APMIS May 2008;116: 400–9
- [57] Bischoff FZ, Heard M, Simpson JL. Somatic DNA alterations in endometriosis: high frequency of chromosome 17 and p53 loss in late-stage endometriosis. J Reprod Immunol May-Jun 2002;55:49–64.
- [58] Nezhat F, Cohen C, Rahaman J, Gretz H, Cole P, Kalir T. Comparative immunohistochemical studies of bcl-2 and p53 proteins in benign and malignant ovarian endometriotic cysts. Cancer Jun 1 2002;94:2935–40.
- [59] Sainz de la Cuesta R, Izquierdo M, Canamero M, Granizo JJ, Manzarbeitia F. Increased prevalence of p53 overexpression from typical endometriosis to atypical endometriosis and ovarian cancer associated with endometriosis. Eur J Obstet Gynecol Reprod Biol Mar 15 2004;113:87–93.
- [60] Akahane T, Sekizawa A, Purwosunu Y, Nagatsuka M, Okai T. The role of p53 mutation in the carcinomas arising from endometriosis. Int J Gynecol Pathol Jul 2007;26:345–51.
- [61] Vercellini P, Trecca D, Oldani S, Fracchiolla NS, Neri A, Crosignani PG. Analysis of p53 and ras gene mutations in endometriosis. Gynecol Obstet Invest 1994;38: 70–1
- [62] Bayramoglu H, Duzcan E. Atypical epithelial changes and mutant p53 gene expression in ovarian endometriosis. Pathol Oncol Res 2001;7:33–8.
- [63] Kurman RJ, Shih I. The origin and pathogenesis of epithelial ovarian cancer: a proposed unifying theory. Am J Surg Pathol Mar 2010;34:433–43.
- [64] Kurman RJ, Shih I. Molecular pathogenesis and extraovarian origin of epithelial ovarian cancer—shifting the paradigm. Hum Pathol Jul 2011;42:918–31.
- [65] Wiegand KC, Shah SP, Al-Agha OM, Zhao Y, Tse K, Zeng T, et al. ARID1A mutations in endometriosis-associated ovarian carcinomas. N Engl J Med Oct 14 2010;363:1532–43.
- [66] Jones S, Wang TL, Shih I, Mao TL, Nakayama K, Roden R, et al. Frequent mutations of chromatin remodeling gene ARID1A in ovarian clear cell carcinoma. Science Oct 8 2010;330:228–31.
- [67] Guan B, Mao TL, Panuganti PK, Kuhn E, Kurman RJ, Maeda D, et al. Mutation and loss of expression of ARID1A in uterine low-grade endometrioid carcinoma. Am J Surg Pathol May 2011;35:625–32.
- [68] Toyokuni S. Role of iron in carcinogenesis: cancer as a ferrotoxic disease. Cancer Sci Jan 2009;100:9–16.
- [69] Kobayashi H, Yamada Y, Kanayama S, Furukawa N, Noguchi T, Haruta S, et al. The role of iron in the pathogenesis of endometriosis. Gynecol Endocrinol Jan 2009;25:39–52.
- [70] Yamaguchi K, Mandai M, Toyokuni S, Hamanishi J, Higuchi T, Takakura K, et al. Contents of endometriotic cysts, especially the high concentration of free iron,

- are a possible cause of carcinogenesis in the cysts through the iron-induced persistent oxidative stress. Clin Cancer Res Ian 1 2008;14:32–40.
- [71] Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? Lancet Feb 17 2001:357:539–45.
- [72] O'Byrne KJ, Dalgleish AG. Chronic immune activation and inflammation as the cause of malignancy. Br J Cancer Aug 17 2001;85:473–83.
- [73] Lebovic DI, Mueller MD, Taylor RN. Immunobiology of endometriosis. Fertil Steril Ian 2001:75:1–10.
- [74] Wu MY, Ho HN. The role of cytokines in endometriosis. Am J Reprod Immunol May 2003;49:285–96.
- [75] Ness RB. Endometriosis and ovarian cancer: thoughts on shared pathophysiology. Am I Obstet Gynecol Jul 2003:189:280–94.
- [76] Darai E, Detchev R, Hugol D, Quang NT. Serum and cyst fluid levels of interleukin (IL) -6, IL-8 and tumour necrosis factor-alpha in women with endometriomas and benign and malignant cystic ovarian tumours. Hum Reprod Aug 2003;18:1681-5.
- [77] Slater M, Cooper M, Murphy CR. Human growth hormone and interleukin-6 are upregulated in endometriosis and endometrioid adenocarcinoma. Acta Histochem 2006;108:13–8.
- [78] Wu MH, Wang CA, Lin CC, Chen LC, Chang WC, Tsai SJ. Distinct regulation of cyclooxygenase-2 by interleukin-1beta in normal and endometriotic stromal cells. J Clin Endocrinol Metab Jan 2005;90:286–95.
- [79] Munkarah AR, Morris R, Baumann P, Deppe G, Malone J, Diamond MP, et al. Effects of prostaglandin E(2) on proliferation and apoptosis of epithelial ovarian cancer cells. J Soc Gynecol Investig May–Jun 2002;9:168–73.
- [80] Murakami M, Kudo I. Recent advances in molecular biology and physiology of the prostaglandin E2-biosynthetic pathway. Prog Lipid Res Jan 2004;43:3–35.
- [81] Rask K, Zhu Y, Wang W, Hedin L, Sundfeldt K. Ovarian epithelial cancer: a role for PGE2-synthesis and signalling in malignant transformation and progression. Mol Cancer Nov 16 2006;5:62.
- [82] Collaborative Group on Hormonal Factors in Breast Cancer. Breast cancer and hormone replacement therapy: collaborative reanalysis of data from 51 epidemiological studies of 52,705 women with breast cancer and 108,411 women without breast cancer. Lancet Oct 11 1997;350:1047–59.
- [83] Beresford SA, Weiss NS, Voigt LF, McKnight B. Risk of endometrial cancer in relation to use of oestrogen combined with cyclic progestagen therapy in postmenopausal women. Lancet Feb 15 1997;349:458–61.
- [84] Morch LS, Lokkegaard E, Andreasen AH, Kruger-Kjaer S, Lidegaard O. Hormone therapy and ovarian cancer. JAMA Jul 15 2009;302:298–305.
- [85] Heaps JM, Nieberg RK, Berek JS. Malignant neoplasms arising in endometriosis. Obstet Gynecol Jun 1990;75:1023–8.
- [86] Oxholm D, Knudsen UB, Kryger-Baggesen N, Ravn P. Postmenopausal endometriosis. Acta Obstet Gynecol Scand Sep 2007;4:1–7.
- [87] Zeitoun KM, Bulun SE. Aromatase: a key molecule in the pathophysiology of endometriosis and a therapeutic target. Fertil Steril Dec 1999;72:961–9.
- [88] Zeitoun K, Takayama K, Sasano H, Suzuki T, Moghrabi N, Andersson S, et al. Deficient 17beta-hydroxysteroid dehydrogenase type 2 expression in endometriosis; failure to metabolize 17beta-estradiol. J Clin Endocrinol Metab Dec 1998;83: 4474–80.
- [89] O'Donnell AJ, Macleod KG, Burns DJ, Smyth JF, Langdon SP. Estrogen receptoralpha mediates gene expression changes and growth response in ovarian cancer cells exposed to estrogen. Endocr Relat Cancer Dec 2005;12:851–66.
- [90] Matsuzaki S, Murakami T, Uehara S, Canis M, Sasano H, Okamura K. Expression of estrogen receptor alpha and beta in peritoneal and ovarian endometriosis. Fertil Steril Jun 2001;75:1198–205.
- [91] Attia GR, Zeitoun K, Edwards D, Johns A, Carr BR, Bulun SE. Progesterone receptor isoform A but not B is expressed in endometriosis. J Clin Endocrinol Metab Aug 2000:85:2897–902.
- [92] Wieser F, Schneeberger C, Tong D, Tempfer C, Huber JC, Wenzl R. PROGINS receptor gene polymorphism is associated with endometriosis. Fertil Steril Feb 2002;77: 309–12.
- [93] Vigano P, Somigliana E, Chiodo I, Abbiati A, Vercellini P. Molecular mechanisms and biological plausibility underlying the malignant transformation of endometriosis: a critical analysis. Hum Reprod Update Jan–Feb 2006;12:77–89.
- [94] Wiegand KC, Lee AF, Al-Agha OM, Chow C, Kalloger SE, Scott DW, et al. Loss of BAF250a (ARID1A) is frequent in high-grade endometrial carcinomas. J Pathol Jul 2011;224:328–33.
- [95] Przybycin CG, Kurman RJ, Ronnett BM, Shih I, Vang R. Are all pelvic (nonuterine) serous carcinomas of tubal origin? Am J Surg Pathol Oct 2010;34:1407–16.